

# Project Notes :

**Project Title:**

**Name: Jessica Froment**

**Note Well:** There are NO SHORT-cuts to reading journal articles and taking notes from them. Comprehension is paramount. You will most likely need to read it several times, so set aside enough time in your schedule.

## Knowledge Gaps :

This list provides a brief overview of the major knowledge gaps for this project, how they were resolved and where to find the information.

Knowledge Gap	Resolved By	Information is located	Date resolved
How do tree networks send info	You tube video	<a href="https://www.youtube.com/watch?v=_tjt8WT5mRs">https://www.youtube.com/watch?v=_tjt8WT5mRs</a>	9/1/24
Difference between arbuscular MN and common MN	Journal Article	<a href="https://nph.onlinelibrary.wiley.com/doi/10.1111/nph.15775">https://nph.onlinelibrary.wiley.com/doi/10.1111/nph.15775</a>	10/1

## Literature Search Parameters:

These searches were performed between (08/14/2024) and 09/27/2024.

List of keywords and databases used during this project.

Database/search engine	Keywords	Summary of search
Brain cancer	Mylin sheaths Brain tumors	Brain cancer is most common within Mylin sheaths as neurons replicate slowly
Mycorrhizal fungi	Tree networks Fungi Comon Mycorrhizal networks Signaling molecules	Plants are able to send signaling molecules through Mycorrhizal fungi. This is pretty understudied and would be a good area of research d

# Tags:

Tag Name	
<p>#RASprotien #microbiology #biology #cancer                      #probability #forecasting #weather # chemistry                      #globalwarming #environment #geneediting                      #mycorrhizal networks</p> <p>#mycorrhizal networks #Carbon #CMNmodel</p> <p>#MN #farming #genetransfer #HGT #DNA                      #bacteria #Symbioses #nutientstransfer #AMF                      #nutrients #Crispr #herbideresistance                      #Glomalin #CO2 #CO2change #N #P #boreal</p>	

## Article #1 Notes: RAS mutations in cancer

Article notes should be on separate sheets

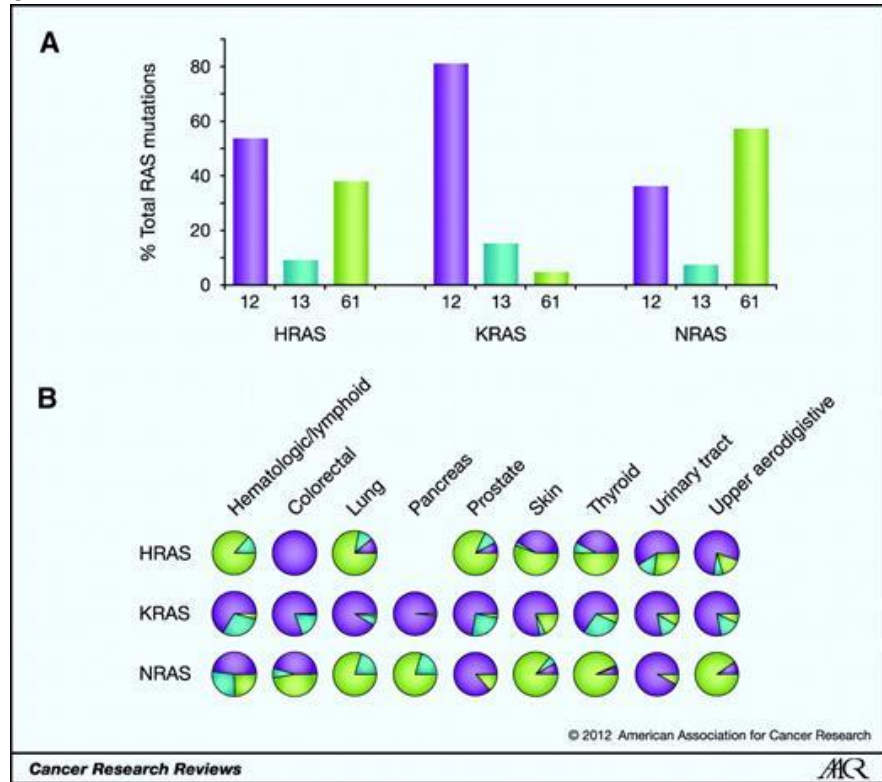
**KEEP THIS BLANK AND USE AS A TEMPLATE**

<b>Source Title</b>	A comprehensive survey of Ras mutations in cancer
<b>Source citation (APA Format)</b>	Prior, I. A., Lewis, P. D., & Mattos, C. (2012). A comprehensive survey of Ras mutations in cancer. <i>Cancer Res</i> 72 (10), 2457–2467. Doi: <a href="https://doi.org/10.1158/0008-5472.CAN-11-2612">https://doi.org/10.1158/0008-5472.CAN-11-2612</a>
<b>Original URL</b>	<a href="https://doi.org/10.1158/0008-5472.CAN-11-2612">https://doi.org/10.1158/0008-5472.CAN-11-2612</a>
<b>Source type</b>	Article
<b>Keywords</b>	RAS protein
<b>#Tags</b>	#RAS protein #microbiology #biology #cancer
<b>Summary of key points + notes (include methodology)</b>	<p>This article from PubMed Central discusses RAS mutations in cancer and how the mutations in isoforms vary among themselves and different types of cancers. The RAS proteins promote cell signaling and growth, and when mutated and activated they promote oncogenesis with unregulated cell division. Understanding RAS genes and proteins is essential towards future methods of battling cancer with cancer research. RAS proteins are encoded with three closely related genes: HRAS, KRAS and NRAS. These proteins are GTP-ases that when activated by GTP create the proteins in signaling pathways that control cell growth. When the GTP binding site is mutated the GTPase-activating proteins (GAP) that inactivated the gene cannot properly bind. This leads to the RAS gene being constantly active, leading to unregulated cell division and creating a tumor.</p> <p>RAS isoforms experience different coupling to specific cancers, as well as the codons and substitutions generating the mutations. For example, KRAS mutations experience 43% G-A mutations at the second base of codon 12 or 13, resulting in G12D or G13D mutations. Almost the rest of the mutations are G-T to produce G12V mutations. NRAS exhibits this at a significantly lower rate. However, HRAS favors mutations at codon 12 to form G12V mutations at a significantly higher rate than both KRAS and NRAS. Also, 80% of all KRAS mutations occur at codon 12, whereas 60% of NRAS are at codon 61, and 50% codon 12 and 40% codon 61 split for HRAS. Even more interestingly are the different percentages of mutations for specific cancers. 90% of pancreatic cancer tumors have KRAS mutations, while hematopoietic tumors are primarily NRAS mutations. One of the reasons for this are the genotoxic agents causing the RAS</p>

mutations. For example, methylnitrosourea (MNU) targets the second base of codon 12 of HRAS and KRAS in many cancers and UV radiation targets pyrimidine dimers resulting in RAS Q61 mutations. The exposure to the particular mutagens results in the specific mutation patterns shown between the isoforms across the different types of cancers. This, however, doesn't explain the differences between RAS isoforms found within the same mutation. There is little experimental analysis on the potential reasons; however, it is speculated from collective data that the differences in the DNA primary sequence, the secondary to quaternary structural effects, and the position of the genes improve or limit the mutagens or repair enzymes access to the isoform. The RAS isoforms also code for different phenotypic responses, supporting the growth of cancer in various ways. KRAS is the most capable of sustaining cancer growth and development, which leads to greater pressure for cancer mutations of this particular isoform.

**Research Question/Problem/ Need** What mutations create cancer, and how do they do so?

**Important Figures** This figure shows the percentages of RAS mutations in different cancers and the percent of where mutations happen in each RAS gene



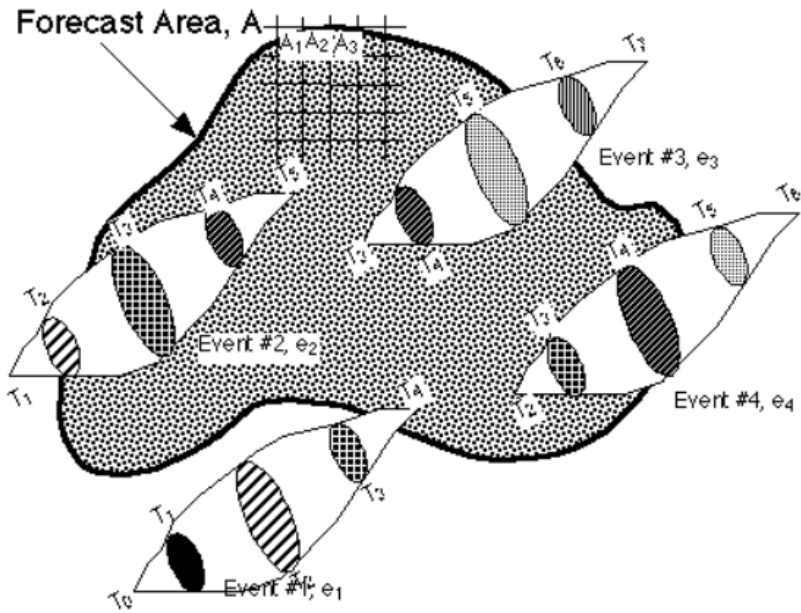
<b>VOCAB: (w/definition)</b>	Isoforms: gene products that are similar but different in function or form
<b>Cited references to follow up on</b>	Gorfe AA, Grant BJ, McCammon JA. Mapping the nucleotide and isoform-dependent structural and dynamical features of Ras proteins. Structure. 2008;16(6):885–96.
<b>Follow up Questions</b>	Can RAS mutations be reversed in order to stop cancer? How do RAS proteins interact in cells? How do RAS proteins vary in structure? What are the different phenotypic responses, and how do they result in different cancers?

## Article #2 Notes: Probability in forecasting

Article notes should be on separate sheets

<b>Source Title</b>	Probabilistic forecasting - A Primer
<b>Source citation (APA Format)</b>	Doswell, C., & Brooks, H. (n.d.). Probabilistic forecasting - A Primer. Doi: <a href="https://www.nssl.noaa.gov/users/brooks/public_html/prob/Probability.html">https://www.nssl.noaa.gov/users/brooks/public_html/prob/Probability.html</a>
<b>Original URL</b>	<a href="https://www.nssl.noaa.gov/users/brooks/public_html/prob/Probability.html">https://www.nssl.noaa.gov/users/brooks/public_html/prob/Probability.html</a>
<b>Source type</b>	Online source
<b>Keywords</b>	Percent of precipitation
<b>#Tags</b>	#probability #forecasting #weather
<b>Summary of key points + notes (include methodology)</b>	This article describes how probability can be used to create polychotomous forecasts, more than two possible outcomes, and accurately predict dangerous weather: severe thunderstorms, tornados,

	<p>etc. The website explains Bayes' theorem and how to forecast mutually related events: one event occurring depends on a certain event happening first. The article also mentions the importance of verifying the predictions made, which proves to be troublesome as it raises a lot of questions due to probabilistic forecasts having significantly more outcomes compared to dichotomous forecasts. Another issue that arrives when forecasting is the area and time period being forecasted. A large area with a large time period will have a near 100% forecast, while a small area and time period will be close to zero. Therefore, it is important to take into consideration the time and area of your forecast. For large amounts it should be attempted to pinpoint areas of high probability within our forecast area and period. If this is impossible, it is best to spread lower probabilities over a wide area. For small amounts climatology frequencies can be used. Climatology frequency uses past events for forecasting when there is little knowledge available. Finally, the article explains how to find the average probability of an event in the area we are forecasting, as events often don't happen in the entirety of the area. This is accomplished by splitting the forecast area into sub-areas, and averaging the probability of the event happening in the area. This also makes it easier to validate the forecast after the event happens. In summary, the article shows how mathematics can be used to predict seemingly unpredictable events, furthering our understanding of how mathematics and the world constantly interact.</p>
<b>Research Question/Problem/ Need</b>	How can we use probability to predict seemingly unpredictable things like nature?

<p><b>Important Figures</b></p>	 <p>Diagram shows how events span across the forecasted area, showing a realistic description of how forecasting looks</p>
<p><b>VOCAB: (w/definition)</b></p>	<p>PoP- percent of precipitation          Polychotomous – more than two possible outcomes          Dichotomous – two outcomes</p>
<p><b>Cited references to follow up on</b></p>	<p>N/A</p>
<p><b>Follow up Questions</b></p>	<p>Can technology, like AI be used to make more accurate forecasts than humans? Can the forecasting reading technology be improved? Can we use forecasting to prevent human casualties before natural disasters?</p>



## Article #3 Notes: the Quantum Origin of the Greenhouse Effect With Carbon Dioxide

Article notes should be on separate sheets

<b>Source Title</b>	Physicists Pinpoint the Quantum Origin of the Greenhouse Effect
<b>Source citation (APA Format)</b>	Howlett, J., Levin, J., Wolchover, N., & Savitsky, Z. (2024b, August 7). <i>Physicists pinpoint the quantum origin of the greenhouse effect</i> . Quanta Magazine. <a href="https://www.quantamagazine.org/physicists-pinpoint-the-quantum-origin-of-the-greenhouse-effect-20240807/">https://www.quantamagazine.org/physicists-pinpoint-the-quantum-origin-of-the-greenhouse-effect-20240807/</a>
<b>Original URL</b>	<a href="https://www.quantamagazine.org/physicists-pinpoint-the-quantum-origin-of-the-greenhouse-effect-20240807/">https://www.quantamagazine.org/physicists-pinpoint-the-quantum-origin-of-the-greenhouse-effect-20240807/</a>
<b>Source type</b>	News Article
<b>Keywords</b>	Carbon dioxide, greenhouse effect
<b>#Tags</b>	#chemistry #globalwarming #environment
<b>Summary of key points + notes (include methodology)</b>	<p>This article talks about how carbon dioxide's quantum structure elevates carbon dioxide's ability to drive the greenhouse effect. The phenomenon in which carbon dioxide traps heat in Earth's atmosphere was discovered by Swedish physicist Svante Arrhenius in 1896. However, the physical reason why carbon dioxide behaves this way would remain unknown until recently.</p> <p>Carbon dioxide can absorb wavelengths slightly shorter or longer than 15 microns, though not as effectively as 15 microns. This range is atypical to other molecules in our atmosphere. When light gets absorbed, the molecule of carbon dioxide can either send it back to Earth or into space. The greenhouse effect is experienced when carbon dioxide sends absorbed light to Earth. Creating a logarithmic scale, as the number of molecules of carbon dioxide doubles, a two to five</p>

	<p>degree temperature rise of the Earth's temperature occurs. Carbon dioxide's unique contribution to global warming is due to its ability to absorb light within a range. A unique phenomenon called Fermi resonance explains that when a photon of 15-micron light is absorbed by carbon dioxide, the light sends the carbon atom swirling at the center point. When this energy of the first motion is doubled, another motion is formed where the two oxygen atoms bob towards and away from the carbon atom. When energy slightly more or less of the first motion is absorbed, hence the importance of carbon dioxide's ability to absorb wavelengths marginally shorter or longer of 15 microns, a new combined motion is created. When carbon dioxide experiences this combined motion, it releases the energy, creating the greenhouse effect. This explains the quantum mechanics behind carbon dioxide's contribution to global warming.</p>
<b>Research Question/Problem/ Need</b>	How does Carbon dioxide contribute to the global warming effect?
<b>Important Figures</b>	N/A
<b>VOCAB: (w/definition)</b>	Microns: unit of measurement used for small molecules
<b>Cited references to follow up on</b>	N/A
<b>Follow up Questions</b>	Can energy be stopped from being absorbed by carbon dioxide? Can carbon dioxide be removed from the environment? Can carbon dioxide be binded with another molecule to prevent the movement mentioned in the article

## Article #4 Notes:

Article notes should be on separate sheets

<b>Source Title</b>	Genome-Editing Technologies: Principles and Applications
<b>Source citation (APA Format)</b>	Prior, I. A., Lewis, P. D., & Mattos, C. (2012). A comprehensive survey of RAS mutations in cancer. <i>Cancer Research</i> , 72(10), 2457–2467. <a href="https://doi.org/10.1158/0008-5472.can-11-2612">https://doi.org/10.1158/0008-5472.can-11-2612</a>
<b>Original URL</b>	<a href="https://doi.org/10.1158/0008-5472.CAN-11-2612">https://doi.org/10.1158/0008-5472.CAN-11-2612</a>
<b>Source type</b>	Journal article
<b>Keywords</b>	Genome
<b>#Tags</b>	#biology #geneediting
<b>Summary of key points + notes (include methodology)</b>	Gene editing, the ability to modify genomic sequences, has expanded scientists' ability to conduct experiments and studies and promoted the possibility of gene therapy. This article studies three technologies that laid the foundation of gene editing: CRISPR-Cas9, TALE nucleases, and zinc-finger nucleases. This article contrasts these three methods and discusses the benefits and disadvantages of using each one. They do this by pulling data from failures and successes of the different types to identify their differences. Overall, CRISPR-Cas9 seemed to be the best as it had the lowest failing rate, best overall success, and the least amount of risk of harming the organism being edited on.
<b>Research Question/Problem/ Need</b>	How are gene editing technologies able to function?
<b>Important Figures</b>	Diagram showing how gene editing works on the molecular level with targeted nucleases

	<p>The diagram illustrates four types of targeted nucleases:</p> <ul style="list-style-type: none"> <li><b>Homing endonuclease:</b> A single protein that binds to a specific DNA sequence and creates a double-strand break.</li> <li><b>ZFN (Zinc-finger Nuclease):</b> Consists of a Zinc-finger DNA-binding domain (1F, 2F, 3F, 4F) and a FokI cleavage domain. Two ZFNs bind to a target sequence, and their FokI domains dimerize to create a double-strand break.</li> <li><b>TALEN (Transcription Activator-Like Effector Nuclease):</b> Consists of a TAL effector DNA-binding domain (with Repeat variable diresidues (RVDs)) and a FokI cleavage domain. Two TALENs bind to a target sequence, and their FokI domains dimerize to create a double-strand break.</li> <li><b>Cas9:</b> A protein that binds to a target DNA sequence adjacent to a PAM (Protospacer Adjacent Motif) sequence, guided by gRNA (guide RNA).</li> </ul> <p><b>Repeat variable diresidues (RVDs) Legend:</b></p> <ul style="list-style-type: none"> <li>NG: T (Red)</li> <li>NI: A (Blue)</li> <li>HD: C (Yellow)</li> <li>NH: G (Green)</li> </ul>
<p><b>VOCAB: (w/definition)</b></p>	<p>Targeted nucleases – allows the editor to target specific areas</p>
<p><b>Cited references to follow up on</b></p>	<p>Burt A. 2003. Site-specific selfish genes as tools for the control and genetic engineering of natural populations. <i>Proc Biol Sci</i> 270: 921–928. [PMC free article] [PubMed] [Google Scholar]</p> <p>Chakraborty S, Ji H, Kabadi AM, Gersbach CA, Christoforou N, Leong KW. 2014. A CRISPR/Cas9-based system for reprogramming cell lineage specification. <i>Stem Cell Rep</i> 3: 940–947. [PMC free article] [PubMed] [Google Scholar]</p>
<p><b>Follow up Questions</b></p>	<p>How can these technologies be expanded upon to limit potential unwanted mutations?</p> <p>Is it ethical to use gene editing on humans even if it prevents deadly diseases?</p> <p>Can gene editing be used to fix mutations in cancerous cells?</p>

## Article #5 Notes: mycorrhizal networks communication

Article notes should be on separate sheets

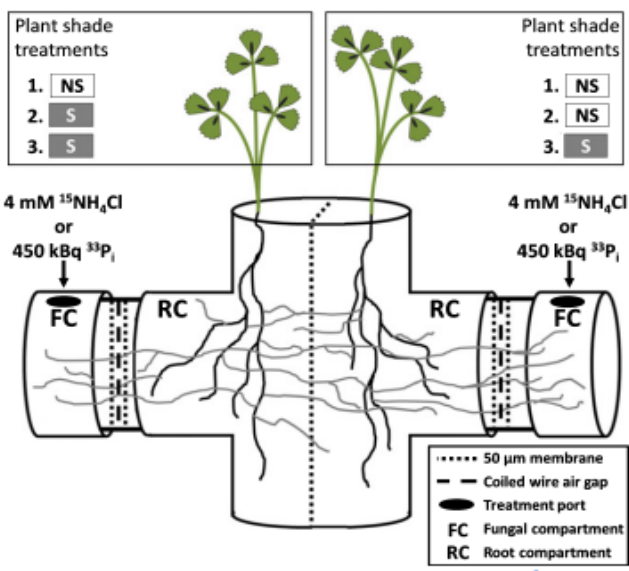
<b>Source Title</b>	Inter-plant communication through mycorrhizal networks mediates complex adaptive behaviour in plant communities
<b>Source citation (APA Format)</b>	Gorzelak, M. A., Asay, A. K., Pickles, B. J., & Gorzelak, S. W. (2015). Inter-plant communication through mycorrhizal networks mediates complex adaptive behaviour in plant communities . <i>AoB PLANTS</i> , 7. <a href="https://doi.org/https://doi.org/10.1093/aobpla/plv050">https://doi.org/https://doi.org/10.1093/aobpla/plv050</a>
<b>Original URL</b>	<a href="https://doi.org/10.1093%2Faobpla%2Fplv050">https://doi.org/10.1093%2Faobpla%2Fplv050</a>
<b>Source type</b>	Journal article
<b>Keywords</b>	Complex adaptive systems, ectomycorrhiza, forests, mycorrhizal networks, plant behaviour, plant communication
<b>#Tags</b>	#mycorrhizal networks
<b>Summary of key points + notes (include methodology)</b>	This article reviews how plants reaction to different stimuli effect the plants within its network. When plants experience stimuli such as nutrient enrichment, stress, etc., it will send out signaling molecules. These signalling molecules are received by connected plants, allowing them to react accordingly. This is important because it shows how plants rely on their relationships with other plants to survive. They did this by comparing data between different previous research to draw large conclusions about these networks. This is done with a fungi called Mycorrhizae, and these fungi have different classes that are preferred. For example, AMF or Arbuscular mycorrhizal fungi is dominate in temperate grasslands, tropical forests and agricultural systems.
<b>Research Question/Problem/ Need</b>	How do environmental stimuli effect signaling in mycorrhizal networks
<b>Important Figures</b>	This diagram shows the different reactions of plants to different stimuli

	<p>This table provides data proving that the plants are reacting to the stimuli received in the network</p>
<p><b>VOCAB: (w/definition)</b></p>	<p>hub trees: central of the forest network          allelochemical: chemical released by an organism that affects the growth and development of other organisms          foliar : relating to leaves</p>
<p><b>Cited references to follow up on</b></p>	<p>Baleshta KE, Simard SW, Guy RD, Chanway CP. 2005. Reducing paper birch density increases Douglas-fir growth rate and Armillaria root disease incidence in southern interior British Columbia. <i>Forest Ecology and Management</i> 208:1–13. 10.1016/j.foreco.2004.07.076          Deslippe JR, Simard SW. 2011. Below-ground carbon transfer among <i>Betula nana</i> may increase with warming in Arctic tundra. <i>New Phytologist</i> 192:689–698. 10.1111/j.1469-8137.2011.03835.x</p>
<p><b>Follow up Questions</b></p>	<p>What are the best ways to model a mycorrhizal network? How do different environmental changes effect the rate at which the networks function? Is it possible for plants to send DNA through these networks?</p>

## Article #6 Notes: Carbon's effect on mycorrhizal networks

Article notes should be on separate sheets

<b>Source Title</b>	<b>Fungal nutrient allocation in common mycorrhizal networks is regulated by the carbon source strength of individual host plants</b>
<b>Source citation (APA Format)</b>	<b>Fellbaum, C., Mensah, J., Cloos, A., Strahan, G., Pfeffer, P., Kier, T., &amp; Beucking, H. (2014). Fungal nutrient allocation in common mycorrhizal networks is regulated by the carbon source strength of individual host plants. <i>New Phytologist</i>, 203, 646–656. doi: 10.1111/nph.12827</b>
<b>Original URL</b>	<a href="https://doi.org/10.1111/nph.12827">10.1111/nph.12827</a>
<b>Source type</b>	<b>Journal article</b>
<b>Keywords</b>	<b>arbuscular mycorrhizal, mutualism, fungus, Carbon</b>
<b>#Tags</b>	<b>#mycorrhizal networks #Carbon #CMNmodel</b>
<b>Summary of key points + notes (include methodology)</b>	This article shows how carbon effects the symbiosis of plant networks. The used a chamber with mesh to prevent roots from moving, and they then gave 33p and 15n and measured their movements. Using isotopic analyses, they found that the result was that the plants with more carbon had more of the isotopes found.

<p><b>Research Question/Problem/Need</b></p>	<p>How does the symbiosis of mycorrhizal networks change as the Plant's Carbon varies?</p>
<p><b>Important Figures</b></p>	 <p><b>Plant shade treatments</b></p> <p>1. NS 2. S 3. S</p> <p>4 mM <math>^{15}\text{NH}_4\text{Cl}</math> or 450 kBq <math>^{33}\text{P}_i</math></p> <p>FC RC RC FC</p> <p>4 mM <math>^{15}\text{NH}_4\text{Cl}</math> or 450 kBq <math>^{33}\text{P}_i</math></p> <p>..... 50 μm membrane --- Coiled wire air gap ● Treatment port FC Fungal compartment RC Root compartment</p> <p><b>Fig. 1</b> The custom-made growth system. A double membrane with an air gap (two sheets of 50-μm nylon mesh divided by a 30-cm-long wire spiral) prevented the diffusion of nutrients from the fungal compartment (FC) to the root compartment (RC), but allowed fungal hyphae to cross from the RCs into the FCs. Three different shade treatments were applied to the plants: (1) both nonshaded (NS/NS), (2) one nonshaded and one shaded (NS/S), and (3) both shaded. To the FCs, 4 mM <math>^{15}\text{NH}_4\text{Cl}</math> or 450 kBq <math>^{33}\text{P}</math>-orthophosphate was added.</p> <p>This is a model that I could potentially use in my project</p>
<p><b>VOCAB: (w/definition)</b></p>	<p><b>Symbiosis:</b> when two plants benefit each other's survival  <b>Ammonium:</b> modified form of ammonia that has an extra hydrogen atom  <b>Putative:</b> generally considered or reputed to be</p>
<p><b>Cited references to follow up on</b></p>	<p>St-Arnaud M, Hamel C, Vimard B, Caron M, Fortin JA. 1996. Enhanced hyphal growth and spore production of the arbuscular mycorrhizal fungus <i>Glomus intraradices</i> in an in vitro system in the absence of host roots. <i>Mycological Research</i> 100: 328–332.  Takanishi I, Ohtomo R, Hayatsu M, Saito M. 2009. Short-chain polyphosphate in arbuscular mycorrhizal roots colonized by <i>Glomus</i> spp.: a possible phosphate pool for host plants. <i>Soil Biology &amp; Biochemistry</i> 41: 1571–1573.  Tanaka Y, Yano K. 2005. Nitrogen delivery to maize via mycorrhizal hyphae depends on the form of N supplied. <i>Plant, Cell &amp; Environment</i> 28: 1247–1254.</p>

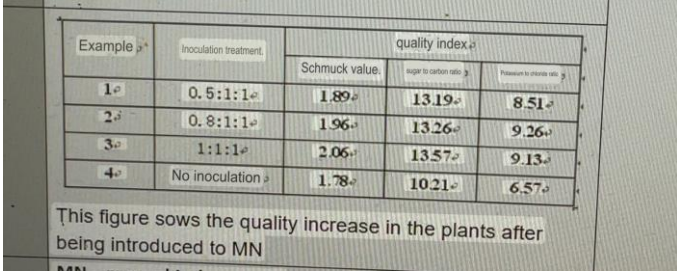


Follow up Questions	How would this change as CO2 in the air changes? What is the rate that this occurs? Is this true for other fungi?
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## Patent #1 Notes: Using Networks in widescale farming.

Article notes should be on separate sheets

Source Title	Application method of arbuscular mycorrhizal fungus in large-scale tobacco cultivation
Source citation (APA Format)	郭涛, 刁向银. (2013). <i>Application method of arbuscular mycorrhizal fungus in large-scale tobacco cultivation</i> (China Patent No. CN103125251B). China Patent Agency. <a href="https://patents.google.com/patent/CN103125251B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi">https://patents.google.com/patent/CN103125251B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi</a>
	<a href="https://patents.google.com/patent/CN103125251B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi">https://patents.google.com/patent/CN103125251B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi</a>
Source type	patent
Keywords	<b>Mycorrhizal networks, tobacco, farming cultivation</b>
#Tags	#MN #farming #biology
Summary of key points + notes (include methodology)	This patent is for a method where they took a MN and used it to improve tobacco to coerce environment adaptability, improve tobacco leaf middle and upper part ratio. The idea is that this would make farming easier and more efficient. They grew the fungi separately, then introduced it to the plants. One network was given a fungicide as the control. When they introduced different diseases, they found the plants with the MN had greater success in their desired categories listed above due to the plants with the MN always having greater nutrients with the diseases than without.

<p><b>Research Question/Problem/Need</b></p>	<p>Can MN be used in widescale farming to improve the cultivation of plants.</p>																												
<p><b>Important Figures</b></p>	<table border="1" data-bbox="378 405 1117 653"> <thead> <tr> <th rowspan="2">实施例</th> <th rowspan="2">接种处理</th> <th colspan="3">品质指标</th> </tr> <tr> <th>施木克值</th> <th>糖碱比</th> <th>钾氮比</th> </tr> </thead> <tbody> <tr> <td>1</td> <td>0.5:1:1</td> <td>1.89</td> <td>13.19</td> <td>8.51</td> </tr> <tr> <td>2</td> <td>0.8:1:1</td> <td>1.96</td> <td>13.26</td> <td>9.26</td> </tr> <tr> <td>3</td> <td>1:1:1</td> <td>2.06</td> <td>13.57</td> <td>9.13</td> </tr> <tr> <td>4</td> <td>不接种菌剂</td> <td>1.78</td> <td>10.21</td> <td>6.57</td> </tr> </tbody> </table> <p data-bbox="1125 642 1393 674">This figure sows the</p> <p data-bbox="367 680 1114 711">quality increase in the plants after being introduced to MN</p>  <p data-bbox="367 993 631 1024">*Chinese translation</p>	实施例	接种处理	品质指标			施木克值	糖碱比	钾氮比	1	0.5:1:1	1.89	13.19	8.51	2	0.8:1:1	1.96	13.26	9.26	3	1:1:1	2.06	13.57	9.13	4	不接种菌剂	1.78	10.21	6.57
实施例	接种处理			品质指标																									
		施木克值	糖碱比	钾氮比																									
1	0.5:1:1	1.89	13.19	8.51																									
2	0.8:1:1	1.96	13.26	9.26																									
3	1:1:1	2.06	13.57	9.13																									
4	不接种菌剂	1.78	10.21	6.57																									
<p><b>VOCAB: (w/definition)</b></p>	<p><b>MN – mycorrhizal networks</b></p> <p><b>Arbuscular mycorrhizal networks – a type of MN where the fungi goes directly into the cell</b></p> <p><b>Inoculation - the action of immunizing someone against a disease by introducing infective material, microorganisms, or vaccine into the body</b></p>																												
<p><b>Cited references to follow up on</b></p>	<p>王发园, 石兆勇, 徐晓锋, 常会庆, 苗艳芳. (2010). <i>Method for reducing heavy metal residues in tobacco by use of arbuscular mycorrhizal fungi</i> (China Patent NO. CN102047808B). China Patent Agency. <a href="https://patents.google.com/patent/CN102047808B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi">https://patents.google.com/patent/CN102047808B/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi</a></p> <p>丁效东, 李淑仪, 詹振寿, 王军. (2012). <i>Method for improving cold and disease resistance of tobacco</i> (China Patent NO. CN102613054A). China Patent Agency. <a href="https://patents.google.com/patent/CN102613054A/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi">https://patents.google.com/patent/CN102613054A/en?q=(Mycorrhizal+fungi)&amp;oq=Mycorrhizal+fungi</a></p>																												

<b>Follow up Questions</b>	<b>Is using MN in farming ethical? Will it not potentially give benefits to weeds? Will this ever be widely implicated with wide fear of GMOs. Can DNA be passed with a AMN</b>
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## Article #7 Notes: Different horizontal gene transfer (HGT) methods plants use

Article notes should be on separate sheets

<b>Source Title</b>	Horizontal gene transfer in the photosphere
<b>Source citation (APA Format)</b>	Van Elsas, J. D., Turner, S., & Bailey, M. J. (2003). Horizontal gene transfer in the phytosphere. <i>New Phytologist</i> , 157(3), 525–537. <a href="https://doi.org/10.1046/j.1469-8137.2003.00697.x">https://doi.org/10.1046/j.1469-8137.2003.00697.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1046/j.1469-8137.2003.00697.x">https://doi.org/10.1046/j.1469-8137.2003.00697.x</a>
<b>Source type</b>	Journal article
<b>Keywords</b>	Gene transfer, HGT, DNA, Bacteria, plant cells

#Tags	#genetransfer #HGT #DNA #biology																																																																							
Summary of key points + notes (include methodology)	This article went over the different ways that plants have been identified to perform HGT. Bacteria are able to form a mating pair formation with Plant cells. This conjugation gene transfer would allow bacteria to intake plant cell DNA, which could then be inserted into another cell. There are different factors that drive HGT: the environment must be good for bacteria colonization and mixing, otherwise the bacteria will be unable to transfer. They used evidence from other experiments to reach this conclusion.																																																																							
Research Question/Problem/ Need	What are the different ways plant cells horizontally transfer DNA?																																																																							
Important Figures	<table border="1" data-bbox="639 814 1289 1360"> <thead> <tr> <th>Species</th> <th>Genomic component</th> <th>Size (Mbp)</th> <th>%GC</th> <th>Accession number</th> </tr> </thead> <tbody> <tr> <td rowspan="3"><i>Mesorhizobium loti</i> MAFF 303099</td> <td>Chromosome</td> <td>7.0</td> <td>62.7<sup>1</sup></td> <td>NC_002678</td> </tr> <tr> <td>pMLa</td> <td>0.35</td> <td>59.3</td> <td>NC_002679</td> </tr> <tr> <td>pMLb</td> <td>0.21</td> <td>59.9</td> <td>NC_002682</td> </tr> <tr> <td rowspan="3"><i>Sinorhizobium meliloti</i> 1021</td> <td>Chromosome</td> <td>3.7</td> <td>62.7</td> <td>NC_003047</td> </tr> <tr> <td>pSymA</td> <td>1.4</td> <td>60.4</td> <td>NC_003037</td> </tr> <tr> <td>pSymB<sup>2</sup></td> <td>1.7</td> <td>62.4</td> <td>NC_003078</td> </tr> <tr> <td rowspan="3"><i>Agrobacterium tumefaciens</i> C58</td> <td>Chromosome</td> <td>2.8</td> <td>59.4</td> <td>NC_003062</td> </tr> <tr> <td>Linear chromosome</td> <td>2.1</td> <td>59.3</td> <td>NC_003063</td> </tr> <tr> <td>pATC58</td> <td>0.54</td> <td>57.3</td> <td>NC_003064</td> </tr> <tr> <td rowspan="3"><i>Brucella melitensis</i></td> <td>pTIC58</td> <td>0.21</td> <td>56.7</td> <td>NC_003065</td> </tr> <tr> <td>Chromosome I</td> <td>2.1</td> <td>57</td> <td>NC_003317</td> </tr> <tr> <td>Chromosome II</td> <td>1.2</td> <td>57</td> <td>NC_003318</td> </tr> <tr> <td rowspan="2"><i>Brucella suis</i></td> <td>Chromosome I</td> <td>2.1</td> <td>57.2</td> <td>NC_004310</td> </tr> <tr> <td>Chromosome II</td> <td>1.2</td> <td>57.3</td> <td>NC_004311</td> </tr> <tr> <td><i>Ralstonia solanearum</i></td> <td>Chromosome</td> <td>3.7</td> <td>67.0</td> <td>NC_003295</td> </tr> </tbody> </table> <p data-bbox="639 1373 1341 1461">Summary of fully sequenced genomes of plant-associated and other bacteria</p>	Species	Genomic component	Size (Mbp)	%GC	Accession number	<i>Mesorhizobium loti</i> MAFF 303099	Chromosome	7.0	62.7 <sup>1</sup>	NC_002678	pMLa	0.35	59.3	NC_002679	pMLb	0.21	59.9	NC_002682	<i>Sinorhizobium meliloti</i> 1021	Chromosome	3.7	62.7	NC_003047	pSymA	1.4	60.4	NC_003037	pSymB <sup>2</sup>	1.7	62.4	NC_003078	<i>Agrobacterium tumefaciens</i> C58	Chromosome	2.8	59.4	NC_003062	Linear chromosome	2.1	59.3	NC_003063	pATC58	0.54	57.3	NC_003064	<i>Brucella melitensis</i>	pTIC58	0.21	56.7	NC_003065	Chromosome I	2.1	57	NC_003317	Chromosome II	1.2	57	NC_003318	<i>Brucella suis</i>	Chromosome I	2.1	57.2	NC_004310	Chromosome II	1.2	57.3	NC_004311	<i>Ralstonia solanearum</i>	Chromosome	3.7	67.0	NC_003295
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VOCAB: (w/definition)	<p data-bbox="639 1577 1003 1608"><b>HGT – horizontal gene transfer</b></p> <p data-bbox="639 1619 1349 1688"><b>Conjugation - the temporary union of two bacteria or unicellular organisms for the exchange of genetic material.</b></p> <p data-bbox="639 1698 1308 1730"><b>enterobacteria - large family of Gram-negative bacteria</b></p>																																																																							
Cited references to follow up on	Bailey MJ, Lilley AK. 2002. Niche colonisation and the dispersal of bacteria and their genes in the natural environment. In: JM																																																																							

	<p>Bullock, RE Kenward, RS Hail, eds. <i>Dispersal ecology</i>. Oxford, UK: Blackwell Publishing Ltd.</p> <p>Bale MJ, Fry JC, Day MJ. 1988. Novel method for studying plasmid transfer in undisturbed river epilithon. <i>Applied and Environmental Microbiology</i> 54: 2756 – 2758.</p>
Follow up Questions	<p>Can the bacteria then transfer that DNA back to a chosen plant cell? How can I control when the transfer happens? How can I control what is being transferred.</p>

## Patent #2 Notes: Different horizontal gene transfer (HGT) methods plants use

Article notes should be on separate sheets

Source Title	<p>USE OF SYNERGISTIC MICROORGANISMS AND NUTRIENTS TO PRODUCE SIGNALS THAT FACILITATE THE GERMINATION AND PLANT ROOT COLONIZATION OF MYCORRHIZAL FUNG IN PHOSPHORUS RICH ENVIRONMENTS</p>
Source citation (APA Format)	<p>Johnson, T. D. (2015). <i>Use of synergistic microorganisms and nutrients to produce signals that facilitate the germination and plant root colonization of mycorrhizal fungi in phosphorus rich environments</i> (U.S. Patent No. US9416061B2). U.S.</p>

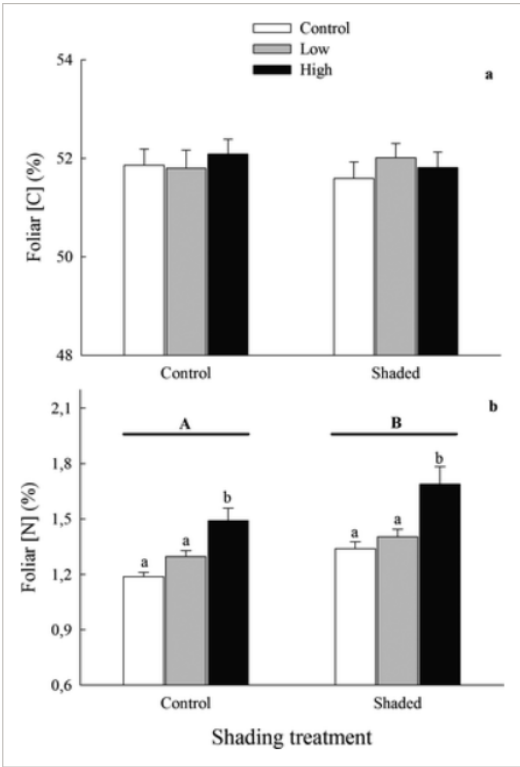
	<p>Patent and Trademark Office.  <a href="https://patents.google.com/patent/US416061B2/en?q=(%22Mycorrhizal+fungi%22)&amp;oq=%22Mycorrhizal+fungi%22">https://patents.google.com/patent/US416061B2/en?q=(%22Mycorrhizal+fungi%22)&amp;oq=%22Mycorrhizal+fungi%22</a></p>																																																																												
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Source type	Patent																																																																												
Keywords	MN, microorganisms, bacteria, phosphorus, nutrients, synergism																																																																												
#Tags	#MN #bacteria #biology																																																																												
Summary of key points + notes (include methodology )	<p>This project focuses on combining synergistic microorganisms to create signals that facilitate germination and plant root colonization of mycorrhizae fungi. The method patented, combining phytate and microorganisms, <i>trichoderma variens</i> fungus, <i>bacillus amyloliquefaciens</i> bacterium, and different mycorrhizae fungi placed so they colonize plant root, creates an effective association of MF with roots in phosphorous rich environments, improving soil aggregation and quality. They tested this invention with different Mycorrhizae propagules, to prove its effectiveness. (figure below) and found that their invention increased plant yield as they plants had more yield with it than without.</p> <p><i>Arbuscular mycorrhizal fungi also have chemotaxic abilities which enable hyphal growth toward the roots of a potential host plant. - important info for project</i></p>																																																																												
Research Question/Problem/ Need	How does the change in mycorrhizal fungi composition affect plant yield?																																																																												
Important Figures	<table border="1"> <thead> <tr> <th>No.</th> <th>Name</th> <th>No Starter</th> <th>Rank</th> <th>10-34-0</th> <th>Rank</th> </tr> </thead> <tbody> <tr> <td>1</td> <td>CHK</td> <td>121.05</td> <td>g</td> <td>7</td> <td>135.37 cde</td> <td>4</td> </tr> <tr> <td>2</td> <td>Myco IF</td> <td>132.72</td> <td>f</td> <td>4</td> <td>133.39 ef</td> <td>5</td> </tr> <tr> <td>3</td> <td>Myco+T.V.+B.A. IF</td> <td>129.22</td> <td>ef</td> <td>6</td> <td>127.77 g</td> <td>7</td> </tr> <tr> <td>4</td> <td>Phytate IF</td> <td>130.96</td> <td>ef</td> <td>5</td> <td>136.24 cde</td> <td>3</td> </tr> <tr> <td>5</td> <td>Phytate+Myco IF</td> <td>137.39</td> <td>cd</td> <td>3</td> <td>132.11 ef</td> <td>6</td> </tr> <tr> <td>6</td> <td>Phytate+T.V.+B.A. IF</td> <td>144.57</td> <td>ab</td> <td>2</td> <td>137.73 bcd</td> <td>2</td> </tr> <tr> <td>7</td> <td>Phytate+Myco+T.V.+B.A. IF</td> <td>147.44</td> <td>a</td> <td>1</td> <td>146.39 a</td> <td>1</td> </tr> <tr> <td colspan="2">LSD (P=.10)</td> <td>4.274</td> <td></td> <td></td> <td>3.475</td> <td></td> </tr> <tr> <td colspan="2">Standard Deviation</td> <td>4.822</td> <td></td> <td></td> <td>3.921</td> <td></td> </tr> <tr> <td colspan="2">CV</td> <td>3.59</td> <td></td> <td></td> <td>2.92</td> <td></td> </tr> </tbody> </table> <p>Shows what mycorrhizal fungi composition creates the biggest yield</p>	No.	Name	No Starter	Rank	10-34-0	Rank	1	CHK	121.05	g	7	135.37 cde	4	2	Myco IF	132.72	f	4	133.39 ef	5	3	Myco+T.V.+B.A. IF	129.22	ef	6	127.77 g	7	4	Phytate IF	130.96	ef	5	136.24 cde	3	5	Phytate+Myco IF	137.39	cd	3	132.11 ef	6	6	Phytate+T.V.+B.A. IF	144.57	ab	2	137.73 bcd	2	7	Phytate+Myco+T.V.+B.A. IF	147.44	a	1	146.39 a	1	LSD (P=.10)		4.274			3.475		Standard Deviation		4.822			3.921		CV		3.59			2.92	
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<b>VOCAB: (w/definition )</b>	Presymbiosis - the development of arbuscular mycorrhizal fungi prior to root colonization Synergistic - the microbial interaction in which both or all the microbial population involved gets benefitted, by supporting each other's growth and proliferation
<b>Cited references to follow up on</b>	Afek et al. Mycorrhizal species.root age-position of Mycorrhizal inoculum influence colonization of cotton.onion, pepper seedlings. J.Amer.Soc. Hort.Sci. 1990.938-942. 115(6).
<b>Follow up Questions</b>	<b>How should I change my mycorrhizal fungi composition to get the best rate of transfer? What mycorrhizal fungi composition would be able to transfer DNA? In what other ways does the mycorrhizal fungi composition affect a plant?</b>

## Article #8 Notes: Nitrogen allocation effects on MN

Article notes should be on separate sheets

<b>Source Title</b>	Greater carbon allocation to mycorrhizal fungi reduces tree nitrogen uptake in a boreal forest
<b>Source citation (APA Format)</b>	Hasselquist, N. J., Metcalfe, D. B., Inselebacher, E., Stangl, Z., Oren, R., Näsholm, T., & Högberg, P. (2016). Greater carbon allocation to mycorrhizal fungi reduces tree nitrogen uptake in a boreal forest. <i>Ecology</i> , 97(4), 1012–1022. <a href="https://doi.org/10.1890/15-1222.1">https://doi.org/10.1890/15-1222.1</a>
<b>Original url</b>	<a href="https://doi.org/10.1890/15-1222.1">https://doi.org/10.1890/15-1222.1</a>

<b>Source type</b>	<b>Journal article</b>																								
<b>Keywords</b>	<b>ectomycorrhizal , symbioses, MN, nutrients transfer, EM</b>																								
<b>#Tags</b>	#MN #Symbioses #nutientstransfer																								
<b>Summary of key points + notes (include methodology)</b>	It is commonly assumed that MN gives N based on the amount of carbon that the host gives to the fungi. This study used large-scale shading treatment to reduce C supply, and gave 3 different levels of N with $^{15}\text{NO}_3^-$ to see how the different N amounts would affect the transfer of N. They found that the amount of Nitrogen given to the MF effected the amount of Nitrogen transferred to the plant, as the isotopic analysis showed a greater amount of N when the N nutrients given was greater.																								
<b>Research Question/Problem/ Need</b>	Does the transfer of nitrogen depend on the amount of carbon and nitrogen available to the MN?																								
<b>Important Figures</b>	 <p>The figure consists of two bar charts, (a) and (b), showing the effect of shading treatment and nitrogen levels on foliar carbon and nitrogen content. Chart (a) shows Foliar C (%) and chart (b) shows Foliar N (%). Both charts compare Control and Shaded treatments, each with three nitrogen levels: Control (white), Low (grey), and High (black). Error bars are present for all data points. Significance letters (a, b) are placed above the bars to indicate statistical differences.</p> <table border="1"> <caption>Data for Figure (a): Foliar C (%)</caption> <thead> <tr> <th>Treatment</th> <th>Control</th> <th>Low</th> <th>High</th> </tr> </thead> <tbody> <tr> <td>Control</td> <td>~51.8</td> <td>~51.8</td> <td>~52.1</td> </tr> <tr> <td>Shaded</td> <td>~51.6</td> <td>~52.0</td> <td>~51.8</td> </tr> </tbody> </table> <table border="1"> <caption>Data for Figure (b): Foliar N (%)</caption> <thead> <tr> <th>Treatment</th> <th>Control</th> <th>Low</th> <th>High</th> </tr> </thead> <tbody> <tr> <td>Control</td> <td>~1.2</td> <td>~1.3</td> <td>~1.5</td> </tr> <tr> <td>Shaded</td> <td>~1.35</td> <td>~1.4</td> <td>~1.65</td> </tr> </tbody> </table>	Treatment	Control	Low	High	Control	~51.8	~51.8	~52.1	Shaded	~51.6	~52.0	~51.8	Treatment	Control	Low	High	Control	~1.2	~1.3	~1.5	Shaded	~1.35	~1.4	~1.65
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<b>VOCAB: (w/definition)</b>	<b>MN – Mycorrhizal network</b> <b>EM - ectomycorrhizal symbioses</b>																								



	<p><b>Ectomycorrhizal symbioses - a ubiquitous plant–fungus interaction in forests, evolved in parallel in fungi</b></p> <p><b>mor-layer - forest humus that forms a layer of largely organic matter distinct from the mineral soil beneath</b></p> <p><b>Perfusate - a fluid used in perfusion.</b></p> <p><b>MF - Mycorrhizal fungi</b></p>
<p><b>Cited references to follow up on</b></p>	<p>Albarracin, M. V., J. Six, B. Z. Houlton, and C. S. Bledsoe. 2013. A nitrogen fertilization field study of carbon-13 and nitrogen-15 transfers in ectomycorrhizas of <i>Pinus sabiniana</i>. <i>Oecologia</i> <b>173</b>: 1439–1450.</p> <p>Bahr, A., M. Ellström, C. Akselsson, A. Ekblad, A. Mikusinska, and H. Wallander. 2013. Growth of ectomycorrhizal fungal mycelium along a Norway spruce forest nitrogen deposition gradient and its effect on nitrogen leakage. <i>Soil Biology &amp; Biochemistry</i> <b>59</b>: 38–48</p> <p>Alberton, O., and T. W. Kuyper. 2009. Ectomycorrhizal fungi associated with <i>Pinus sylvestris</i> seedlings respond differently to increased carbon and nitrogen availability: implications for ecosystem responses to global change. <i>Global Change Biology</i> 15: 166–175.</p>
<p><b>Follow up Questions</b></p>	<p><b>Would this explain the insignificantly different data found in the article ‘Fungal nutrient allocation in common mycorrhizal networks is regulated by the carbon source strength of individual host plants’? How would this work for other nutrients? Does the amount of nitrogen effect other plants in the network?</b></p>

## Article #9 Notes: Using MN networks in farming

Article notes should be on separate sheets

<b>Source Title</b>	Is there a role for arbuscular mycorrhizal fungi in production agriculture?
<b>Source citation (APA Format)</b>	Ryan, M. H., & Graham, J. H. (2002). Is there a role for arbuscular mycorrhizal fungi in production agriculture? <i>Plant and Soil</i> , 244(1/2), 263–271. <a href="https://doi.org/10.1023/a:1020207631893">https://doi.org/10.1023/a:1020207631893</a>
<b>Original url</b>	<a href="https://doi.org/10.1023/a:1020207631893">https://doi.org/10.1023/a:1020207631893</a>
<b>Source type</b>	Journal article

<b>Keywords</b>	<b>AMF, plant growth, nutrients, AM colonisation, farming</b>																																																																																																		
<b>#Tags</b>	#AMF #MN #farming #nutrients																																																																																																		
<b>Summary of key points + notes (include methodology)</b>	This article argues that AMF does not greatly affect nutrition and growth of plants in production-orientated agricultural systems. They did this by taking different studies drawn that have proven high AMF effect, and stated different reasons that would have lead to this conclusion. They then went over the structure of AMF and why it wouldn't work within the context of the experiment, and they generally hypothesized that nutrients levels effect nutrients intake much more than AMF. They also generally hypothesized that AMF effects are being over estimated due to use of field-based case studies, which would result in environment effecting results. They concluded that while AMF effect nutrients and growth, it is not to the degree that has been previously though.																																																																																																		
<b>Research Question/Problem/Need</b>	Does arbuscular mycorrhizal fungi effect the nutrition and growth of plants in production-orientated agricultural systems?																																																																																																		
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<b>VOCAB: (w/definition)</b>	<b>AMF</b> - arbuscular mycorrhizal fungi CMN – common mycorrhizal fungi dazomet fumigant - soil fumigant that acts as a herbicide, fungicide, slimicide, and nematicide Colonization- fungi growth																																																																																																		
<b>Cited references to follow up on</b>	Thompson J P 1987 Decline of vesicular-arbuscular mycorrhizas in long fallow disorder of field crops and its expression in phosphorus deficiency in sunflower. Aust. J. Agric. Res. 38, 847–867.																																																																																																		

	<p>Thingstrup I, Rubæk G, Sibbesen E and Jakobsen I 1998 Flax (<i>Linum usitatissimum</i> L.) depends on arbuscular mycorrhizal fungi for growth and P uptake at intermediate but not high soil P levels in the field. <i>Plant Soil</i> 203, 37–46.</p> <p>Miller R M and Jastrow J D 2000 Mycorrhizal fungi influence soil structure. In <i>Arbuscular Mycorrhizas: Physiology and Function</i>. Eds. Y Kapulnik and D D Douds. pp. 3–18. Kluwer, Dordrecht.</p>
<b>Follow up Questions</b>	<b>Are there any studies proving this to be true via experiments? If true, how will this effect my experiment? Is this also true of CMN?</b>

## Article #10 Notes: CRISPR use in plants

Article notes should be on separate sheets

<b>Source Title</b>	Application of CRISPR/Cas9-mediated gene editing for the development of herbicide-resistant plants
<b>Source citation (APA Format)</b>	Han, Y.-J., & Kim, J.-I. (2019). Application of CRISPR/Cas9-mediated gene editing for the development of herbicide-resistant plants. <i>Plant Biotechnology Reports</i> , 13(5), 447–457. <a href="https://doi.org/10.1007/s11816-019-00575-8">https://doi.org/10.1007/s11816-019-00575-8</a>
<b>Original url</b>	<a href="https://doi.org/10.1007/s11816-019-00575-8">https://doi.org/10.1007/s11816-019-00575-8</a>
<b>Source type</b>	Journal article
<b>Keywords</b>	<b>CRISPR/Cas9</b> <b>Gene editing</b> <b>Genome engineering</b> <b>Herbicide tolerance</b> <b>Crop improvement</b>
<b>#Tags</b>	#geneediting #Crispr #farming #herbideresistance
<b>Summary of key points + notes (include methodology)</b>	<p>This article went through the different benefits of using genetically engineering. By genetically engineering farm plants to be resistant to the herbicide, it allows farmers to kill the weeds while allowing the plants to live. This article hypothesized several potential gene editing, CESA3 and SF3B1, that could be used to kill weeds. They tested this out by editing these genes in the plants (to make them HR) and then giving a non edited and an edited one herbicides. They found their hypothesis to be proven to be correct as the HR plants survived while the weeds died.</p>

<b>Research Question/Problem/Need</b>	Can herbicide resistance be used to increase farming productivity?																																																
<b>Important Figures</b>	<table border="1"> <thead> <tr> <th>Herbicide</th> <th>Gene</th> <th>Gene product</th> <th>Crop</th> </tr> </thead> <tbody> <tr> <td>2,4-D<sup>b</sup></td> <td>AAO-1</td> <td>Aryloxyalkanoate dioxygenase-1 from <i>Delftia acidovorans</i></td> <td>Maize, soybean</td> </tr> <tr> <td>Dicamba<sup>c</sup></td> <td>DMO</td> <td>Dicamba monooxygenase from <i>Stenotrophomonas maltophilia</i> strain DI-6</td> <td>Soybean</td> </tr> <tr> <td rowspan="2">Glufosinate</td> <td>BAR</td> <td>Bialaphos resistance gene from <i>Streptomyces hygroscopicus</i></td> <td>Canola, chicory, cotton, maize, rice, soybean</td> </tr> <tr> <td>PAT</td> <td>Phosphinothricin N-acetyltransferase from <i>Streptomyces viridochromogenes</i></td> <td>Canola, cotton, maize, soybean, sugar beet</td> </tr> <tr> <td>Oxynil</td> <td>BXN</td> <td>Bromoxynil nitrilase from <i>Klebsiella pneumoniae</i> subsp. <i>ozaenae</i></td> <td>Canola, cotton, tobacco</td> </tr> <tr> <td rowspan="4">Glyphosate</td> <td>GAT4601</td> <td>Glyphosate N-acetyltransferase from <i>Bacillus licheniformis</i></td> <td>Canola, Maize</td> </tr> <tr> <td>CP4EPSPS</td> <td>An herbicide tolerant form of 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) from <i>Agrobacterium tumefaciens</i> strain CP4</td> <td>Alfalfa, canola, cotton, creeping bentgrass, maize, potato, soybean, sugar beet, wheat</td> </tr> <tr> <td>2MEPSPS</td> <td>A double mutant version (T102I/P106S) of EPSPS from <i>Zea mays</i></td> <td>Cotton, maize, soybean</td> </tr> <tr> <td>MEPSPS</td> <td>A modified EPSPS (two amino acid substitutions) from <i>Zea mays</i></td> <td>Maize</td> </tr> <tr> <td>Isoxaflutole</td> <td>HPPDPF W336</td> <td>A modified (G336W) p-hydroxyphenylpyruvate dioxygenase (HPPD) from <i>Pseudomonas fluorescens</i> strain A32</td> <td>Cotton, soybean</td> </tr> <tr> <td>Mesotrione</td> <td>AvHPPD-03</td> <td>An HPPD isozyme (missing A111) from <i>Avena sativa</i> (AvHPPD)</td> <td>Soybean</td> </tr> <tr> <td>Sulfonylurea</td> <td>SURB</td> <td>A mutant (P196A/W573L) of Acetolactate synthase (ALS) on locus SuRB from <i>Nicotiana tabacum</i></td> <td>Carnation</td> </tr> </tbody> </table> <p>Currently known HR plants</p>	Herbicide	Gene	Gene product	Crop	2,4-D <sup>b</sup>	AAO-1	Aryloxyalkanoate dioxygenase-1 from <i>Delftia acidovorans</i>	Maize, soybean	Dicamba <sup>c</sup>	DMO	Dicamba monooxygenase from <i>Stenotrophomonas maltophilia</i> strain DI-6	Soybean	Glufosinate	BAR	Bialaphos resistance gene from <i>Streptomyces hygroscopicus</i>	Canola, chicory, cotton, maize, rice, soybean	PAT	Phosphinothricin N-acetyltransferase from <i>Streptomyces viridochromogenes</i>	Canola, cotton, maize, soybean, sugar beet	Oxynil	BXN	Bromoxynil nitrilase from <i>Klebsiella pneumoniae</i> subsp. <i>ozaenae</i>	Canola, cotton, tobacco	Glyphosate	GAT4601	Glyphosate N-acetyltransferase from <i>Bacillus licheniformis</i>	Canola, Maize	CP4EPSPS	An herbicide tolerant form of 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) from <i>Agrobacterium tumefaciens</i> strain CP4	Alfalfa, canola, cotton, creeping bentgrass, maize, potato, soybean, sugar beet, wheat	2MEPSPS	A double mutant version (T102I/P106S) of EPSPS from <i>Zea mays</i>	Cotton, maize, soybean	MEPSPS	A modified EPSPS (two amino acid substitutions) from <i>Zea mays</i>	Maize	Isoxaflutole	HPPDPF W336	A modified (G336W) p-hydroxyphenylpyruvate dioxygenase (HPPD) from <i>Pseudomonas fluorescens</i> strain A32	Cotton, soybean	Mesotrione	AvHPPD-03	An HPPD isozyme (missing A111) from <i>Avena sativa</i> (AvHPPD)	Soybean	Sulfonylurea	SURB	A mutant (P196A/W573L) of Acetolactate synthase (ALS) on locus SuRB from <i>Nicotiana tabacum</i>	Carnation
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<b>VOCAB: (w/definition)</b>	<b>HR- herbicide-resistant deregulation - the removal of regulations or restrictions, especially in a particular industry.</b>																																																
<b>Cited references to follow up on</b>	<p>Ma X, Zhu Q, Chen Y, Liu YG (2016) CRISPR/Cas9 platforms for genome editing in plants: developments and applications. <i>Mol Plant</i> 9:961–974</p> <p>Li J, Meng XB, Zong Y, Chen KL, Zhang HW, Liu JX, Li JY, Gao CX (2016) Gene replacements and insertions in rice by intron targeting using CRISPR-Cas9. <i>Nat Plants</i> 2:16139</p> <p>Li Z, Liu ZB, Xing A, Moon BP, Koellhoffer JP, Huang L, Ward RT, Clifton E, Falco SC, Cigan AM (2015) Cas9-guide RNA directed genome editing in soybean. <i>Plant Physiol</i> 169:960–970</p>																																																
<b>Follow up Questions</b>	<b>What are the potential risks of using gene editing? Would people want to use GMO's? Could I relate my project to farming benefits?</b>																																																

# Article #11 Notes: Mycorrhizal responses to nitrogen fertilization in boreal ecosystems

Article notes should be on separate sheets

<b>Source Title</b>	Mycorrhizal responses to nitrogen fertilization in boreal ecosystems: potential consequences for soil carbon storage
<b>Source citation (APA Format)</b>	Treseder, K. K., Turner, K. M., & Mack, M. C. (2006). Mycorrhizal responses to nitrogen fertilization in boreal ecosystems: Potential consequences for Soil Carbon Storage. <i>Global Change Biology</i> , 13(1), 78–88. <a href="https://doi.org/10.1111/j.1365-2486.2006.01279.x">https://doi.org/10.1111/j.1365-2486.2006.01279.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1111/j.1365-2486.2006.01279.x">https://doi.org/10.1111/j.1365-2486.2006.01279.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	<b>Mycorrhizal Fungi</b> <b>Nitrogen Fertilization</b> <b>Boreal Ecosystems</b> <b>Soil Carbon Storage</b> <b>Carbon Cycling</b> <b>Arbuscular Mycorrhizal Fungi (AMF)</b>
<b>#Tags</b>	#AMF #MN #boreal #nutrients #moleculertrasfer
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- They used several boreal forest sites on Alaska and treated them to N fertilization, some left without as controls</li> <li>- Findings <ul style="list-style-type: none"> <li>o Increase glomalin levels lead to reduction of 50 g C m<sup>-2</sup> in total mycorrhizal carbon pools</li> </ul> </li> </ul>

	<ul style="list-style-type: none"> <li>○ Increased the abundance of root AM structures across all sites, with large increase ECM structures, however no effect on soil AM hyphae</li> <li>○ Carbon sequestering was modest compared to that in glomalin --&gt; glomalin significant in soil carbon storage</li> </ul>
<p><b>Research Question/Problem/Need</b></p>	<p>How will the abundance of mycorrhizal fungi and glomalin change under N fertilization in boreal ecosystems.</p>
<p><b>Important Figures</b></p>	<p>Shows the length of the hyphal as the time goes on (<b>set up</b>)</p>
<p><b>VOCAB: (w/definition)</b></p>	<p>hyphal - filamentous structures that make up fungi          glomalin - glycoprotein, or sugar-protein compound, that's produced by arbuscular mycorrhizal fungi in soil and plant roots          boreal ecosystems - a large, northern forest biome</p>
<p><b>Cited references to follow up on</b></p>	<p>Driver, J. D., Holben, W. E., &amp; Rillig, M. C. (2005). Characterization of glomalin as a hyphal wall component of arbuscular mycorrhizal fungi. <i>Soil Biology and</i></p>

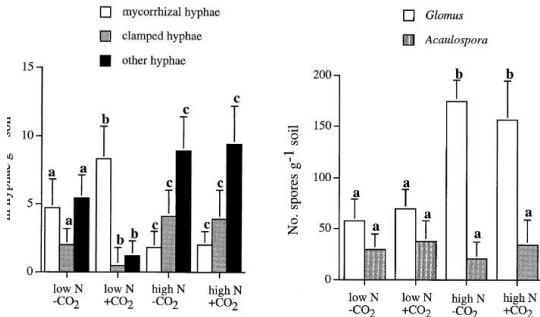


	<p><i>Biochemistry</i>, 37(1), 101–106.  <a href="https://doi.org/10.1016/j.soilbio.2004.06.011">https://doi.org/10.1016/j.soilbio.2004.06.011</a></p> <p>Friese, C. F., &amp; Allen, M. F. (1991). The spread of VA mycorrhizal fungal hyphae in the soil: Inoculum types and external hyphal architecture. <i>Mycologia</i>, 83(4), 409–418.  <a href="https://doi.org/10.1080/00275514.1991.12026030">https://doi.org/10.1080/00275514.1991.12026030</a></p> <p>Klironomos, J., Rillig, M., Allen, M., Zak, D., Kubiske, M., &amp; Pregitzer, K. (1997a). Soil fungal-arthropod responses to populus tremuloides grown under enriched atmospheric CO<sub>2</sub> under field conditions. <i>Global Change Biology</i>, 3(6), 473–478. <a href="https://doi.org/10.1046/j.1365-2486.1997.00085.x">https://doi.org/10.1046/j.1365-2486.1997.00085.x</a></p>
<b>Follow up Questions</b>	<p><b>How would Glomalin work on plants not within a boreal forest? Would nitrogen fertilization improve this with all types of AM fungi? Was there any change in the structures of the fungi across the stages or site conditions</b></p>

## Article #12 Notes: Populus tremuloides with CO<sub>2</sub> under field conditions

Article notes should be on separate sheets

<b>Source Title</b>	Soil fungal-arthropod responses to Populus tremuloides grown under enriched atmospheric CO <sub>2</sub> under field conditions
<b>Source citation (APA Format)</b>	<p>Klironomos, J., Rillig, M., Allen, M., Zak, D., Kubiske, M., &amp; Pregitzer, K. (1997a). Soil fungal-arthropod responses to populus tremuloides grown under enriched atmospheric CO<sub>2</sub> under field conditions. <i>Global Change Biology</i>, 3(6), 473–478. <a href="https://doi.org/10.1046/j.1365-2486.1997.00085.x">https://doi.org/10.1046/j.1365-2486.1997.00085.x</a></p>

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Source type	Article																														
Keywords	<b>Soil Fungi</b> <b>Arthropod Communities</b> <b>Elevated CO<sub>2</sub></b> <b>Field Conditions</b> <b>Fungal Hyphae</b> <b>Soil Respiration</b>																														
#Tags	#mycorrhizal networks #Carbon #CMNmodel																														
Summary of key points + notes (include methodology)	<ul style="list-style-type: none"> <li>- Took populus tremuloides and grew under two conditions ambient CO<sub>2</sub> and elevated CO<sub>2</sub>. They then put in two different N treatments (limited N and enriched N) into the soil. They measured nutrient levels as well as plant and fungi growth.</li> <li>- Findings <ul style="list-style-type: none"> <li>○ Low N <ul style="list-style-type: none"> <li>▪ Elevated CO<sub>2</sub> increased hyphal length by 77% for AM fungi</li> </ul> </li> <li>○ High N <ul style="list-style-type: none"> <li>▪ Reduced AMF by 71%, but more research found that they opposite relationship was true for non mycorrhizal fungi</li> </ul> </li> </ul> </li> </ul>																														
Research Question/Problem/Need	What is the influence of elevated CO <sub>2</sub> and soil N availability on the growth of arbuscular mycorrhizal and non-mycorrhizal fungi, and on the number of mycophagous soil microarthropods associated with the roots of <i>Populus tremuloides</i> ?																														
Important Figures	 <p>The figure consists of two bar charts. The left chart shows 'hyphal length (mm g<sup>-1</sup> root)' on the y-axis (0 to 15) for four conditions: low N -CO<sub>2</sub>, low N +CO<sub>2</sub>, high N -CO<sub>2</sub>, and high N +CO<sub>2</sub>. The legend indicates three types of hyphae: mycorrhizal (white), clamped (grey), and other (black). The right chart shows 'No. spores g<sup>-1</sup> soil' on the y-axis (0 to 200) for the same four conditions, with a legend for <i>Glomus</i> (white) and <i>Acaulospora</i> (grey). Error bars and significance letters (a, b, c) are present on all bars.</p> <table border="1"> <caption>Approximate data from the figure</caption> <thead> <tr> <th>Condition</th> <th>Mycorrhizal hyphae (mm g<sup>-1</sup> root)</th> <th>Clamped hyphae (mm g<sup>-1</sup> root)</th> <th>Other hyphae (mm g<sup>-1</sup> root)</th> <th><i>Glomus</i> spores (g<sup>-1</sup> soil)</th> <th><i>Acaulospora</i> spores (g<sup>-1</sup> soil)</th> </tr> </thead> <tbody> <tr> <td>low N -CO<sub>2</sub></td> <td>~5.5 (a)</td> <td>~2.5 (a)</td> <td>~5.5 (a)</td> <td>~55 (a)</td> <td>~30 (a)</td> </tr> <tr> <td>low N +CO<sub>2</sub></td> <td>~8.5 (b)</td> <td>~2.5 (a)</td> <td>~1.5 (b)</td> <td>~70 (a)</td> <td>~40 (a)</td> </tr> <tr> <td>high N -CO<sub>2</sub></td> <td>~3.5 (c)</td> <td>~4.5 (c)</td> <td>~9.5 (c)</td> <td>~20 (a)</td> <td>~35 (a)</td> </tr> <tr> <td>high N +CO<sub>2</sub></td> <td>~2.5 (c)</td> <td>~4.5 (c)</td> <td>~9.5 (c)</td> <td>~160 (b)</td> <td>~35 (a)</td> </tr> </tbody> </table>	Condition	Mycorrhizal hyphae (mm g <sup>-1</sup> root)	Clamped hyphae (mm g <sup>-1</sup> root)	Other hyphae (mm g <sup>-1</sup> root)	<i>Glomus</i> spores (g <sup>-1</sup> soil)	<i>Acaulospora</i> spores (g <sup>-1</sup> soil)	low N -CO <sub>2</sub>	~5.5 (a)	~2.5 (a)	~5.5 (a)	~55 (a)	~30 (a)	low N +CO <sub>2</sub>	~8.5 (b)	~2.5 (a)	~1.5 (b)	~70 (a)	~40 (a)	high N -CO <sub>2</sub>	~3.5 (c)	~4.5 (c)	~9.5 (c)	~20 (a)	~35 (a)	high N +CO <sub>2</sub>	~2.5 (c)	~4.5 (c)	~9.5 (c)	~160 (b)	~35 (a)
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	Effect of CO <sub>2</sub> and N amount on the hyphae length of fungi
<b>VOCAB: (w/definition)</b>	<ul style="list-style-type: none"> <li>- Arbuscular Mycorrhizal Fungi: A type of symbiotic fungi that form associations with plant roots, facilitating nutrient exchange (e.g., phosphorus) and receiving carbon from the host plant.</li> <li>- Hyphal Length: A measure of the total length of fungal filaments (hyphae) in a given volume of soil, reflecting fungal growth and activity.</li> <li>- Priming Effect: A phenomenon where increased microbial or fungal activity accelerates the decomposition of soil organic matter, potentially releasing stored carbon.</li> </ul>
<b>Cited references to follow up on</b>	<p>Oves, M., Hussain, F. M., Ismail, I. M. I., Felemban, N. M., &amp; Qari, H. A. (2017). Microbiological carbon sequestration. <i>Advances in Environmental Engineering and Green Technologies</i>, 108–133. <a href="https://doi.org/10.4018/978-1-5225-2325-3.ch005">https://doi.org/10.4018/978-1-5225-2325-3.ch005</a></p> <p>Bonito, G., Hameed, K., Ventura, R., Krishnan, J., Schadt, C. W., &amp; Vilgalys, R. (2016). Isolating a functionally relevant guild of fungi from the root microbiome of populus. <i>Fungal Ecology</i>, 22, 35–42. <a href="https://doi.org/10.1016/j.funeco.2016.04.007">https://doi.org/10.1016/j.funeco.2016.04.007</a></p> <p>Reddy, P. P. (2014). Impacts on plant pathogens. <i>Climate Resilient Agriculture for Ensuring Food Security</i>, 151–177. <a href="https://doi.org/10.1007/978-81-322-2199-9_8">https://doi.org/10.1007/978-81-322-2199-9_8</a></p>
<b>Follow up Questions</b>	<b>Why did AM increase hyphal length under low N and high CO<sub>2</sub>? What drives this relationship? How does the reduction of the hyphal length affect the rest of the system? How does it affect the livelihood of the fungi?</b>

Article #13 Notes: Mycorrhizae with quantity of carbon allocated below ground changing

Article notes should be on separate sheets

<b>Source Title</b>	Mycorrhizae alter quality and quantity of carbon allocated below ground
<b>Source citation (APA Format)</b>	Rygiewicz, P. T., & Andersen, C. P. (1994). Mycorrhizae alter quality and quantity of carbon allocated below ground. <i>Nature</i> , 369(6475), 58–60. <a href="https://doi.org/10.1038/369058a0">https://doi.org/10.1038/369058a0</a>
<b>Original url</b>	<a href="https://doi.org/10.1038/369058a0">https://doi.org/10.1038/369058a0</a>
<b>Source type</b>	Article
<b>Keywords</b>	<b>Mycorrhizae</b> <b>Carbon Allocation</b> <b>Below-Ground Carbon</b> <b>Fungal Hyphae</b> <b>Carbon Quality</b> <b>Root-Fungal Symbiosis</b> <b>Soil Organic Carbon</b> <b>Glomalin</b>
<b>#Tags</b>	#mycorrhizal networks #Carbon #CMNmodel #Glomalin
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- Used colonized and non-colonized plants exposed to CO<sub>2</sub> with <sup>13</sup>C and then measured the carbon in above ground biomass and below ground</li> <li>- Findings <ul style="list-style-type: none"> <li>○ Plants have more C allocation to below ground biomass (especially the roots and fungal hyphae) <ul style="list-style-type: none"> <li>▪ Leads to increase below ground respiration and reduction of C above ground</li> </ul> </li> </ul> </li> </ul>

	<ul style="list-style-type: none"> <li>○ Carbon is directed towards carbon pools with rapid turnover rates (effects retention time C in soil)</li> <li>○ Mycorrhizal influence quantity of carbon allocation below ground impacting soil carbon storage</li> <li>○ Mycorrhizae are influenced by CO<sub>2</sub></li> </ul>																																																																						
<p><b>Research Question/Problem/Need</b></p>	<p>How do mycorrhizal associations influence distribution of C allocated to Mycorrhizae?</p>																																																																						
<p><b>Important Figures</b></p>	<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: left;">Fraction</th> <th style="text-align: center;">Non-inoculated</th> <th style="text-align: center;">Ectomyc.</th> <th style="text-align: center;">Non-inoculated</th> <th style="text-align: center;">Ectomyc.</th> </tr> <tr> <th></th> <th colspan="2" style="text-align: center;">Dry weight (mg)</th> <th colspan="2" style="text-align: center;">Relative activity (Bq <sup>14</sup>C mg<sup>-1</sup>)</th> </tr> </thead> <tbody> <tr> <td>Total plant</td> <td style="text-align: center;">2,080</td> <td style="text-align: center;">1,878*</td> <td></td> <td></td> </tr> <tr> <td>Above ground</td> <td></td> <td></td> <td></td> <td></td> </tr> <tr> <td>Total</td> <td style="text-align: center;">940</td> <td style="text-align: center;">868*</td> <td style="text-align: center;">50.0</td> <td style="text-align: center;">44.0</td> </tr> <tr> <td>Bud and stem</td> <td style="text-align: center;">266</td> <td style="text-align: center;">238</td> <td style="text-align: center;">20.0</td> <td style="text-align: center;">22.4</td> </tr> <tr> <td>Needles</td> <td style="text-align: center;">674</td> <td style="text-align: center;">630</td> <td style="text-align: center;">27.0</td> <td style="text-align: center;">27.0</td> </tr> <tr> <td>Below ground</td> <td></td> <td></td> <td></td> <td></td> </tr> <tr> <td>Total</td> <td style="text-align: center;">1,137</td> <td style="text-align: center;">1,010</td> <td style="text-align: center;">15.3</td> <td style="text-align: center;">22.1**</td> </tr> <tr> <td>Roots (host)</td> <td style="text-align: center;">1,137</td> <td style="text-align: center;">910***</td> <td style="text-align: center;">15.3</td> <td style="text-align: center;">19.8</td> </tr> <tr> <td>Active hyphae†</td> <td style="text-align: center;">n.p.</td> <td style="text-align: center;">100***</td> <td style="text-align: center;">n.p.</td> <td style="text-align: center;">40.1***</td> </tr> <tr> <td>Coarse roots</td> <td style="text-align: center;">936</td> <td style="text-align: center;">741**</td> <td style="text-align: center;">14.6</td> <td style="text-align: center;">17.7</td> </tr> <tr> <td>Fine roots (host)</td> <td style="text-align: center;">201</td> <td style="text-align: center;">169</td> <td style="text-align: center;">16.9</td> <td style="text-align: center;">23.4*</td> </tr> <tr> <td>Active hyphae</td> <td style="text-align: center;">n.p.</td> <td style="text-align: center;">100***</td> <td style="text-align: center;">n.p.</td> <td style="text-align: center;">32.8***</td> </tr> </tbody> </table> <p style="font-size: small;">Values are means of numbers that were calculated using individual seedling data of <sup>14</sup>C allocation to fractions and the respective fraction dry weight. The individual seedling relative activity value was then standardized to 10<sup>9</sup> Bq total radioisotope assimilation by needles. All statistical conventions are as in Fig. 2. n.p., Not present.  † See Fig. 2 legend for estimates of hyphal mass in ectomycorrhizal root tips.</p> <p>Fraction of dry weights and activity fractions based on carbon in the needles</p>	Fraction	Non-inoculated	Ectomyc.	Non-inoculated	Ectomyc.		Dry weight (mg)		Relative activity (Bq <sup>14</sup> C mg <sup>-1</sup> )		Total plant	2,080	1,878*			Above ground					Total	940	868*	50.0	44.0	Bud and stem	266	238	20.0	22.4	Needles	674	630	27.0	27.0	Below ground					Total	1,137	1,010	15.3	22.1**	Roots (host)	1,137	910***	15.3	19.8	Active hyphae†	n.p.	100***	n.p.	40.1***	Coarse roots	936	741**	14.6	17.7	Fine roots (host)	201	169	16.9	23.4*	Active hyphae	n.p.	100***	n.p.	32.8***
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<p><b>VOCAB: (w/definition)</b></p>	<p>Hyphae - each of the branching filaments that make up the mycelium of a fungus.</p> <p>Allocation - the action or process of distributing something.</p> <p>Assimilation - movement of digested food into cells</p> <p>Respiration - chemical process that releases energy from food</p>																																																																						
<p><b>Cited references to follow up on</b></p>	<p>Dixon, R. K., Solomon, A. M., Brown, S., Houghton, R. A., Trexler, M. C., &amp; Wisniewski, J. (1994). Carbon Pools and flux of global forest ecosystems. <i>Science</i>, 263(5144), 185–190.  <a href="https://doi.org/10.1126/science.263.5144.185">https://doi.org/10.1126/science.263.5144.185</a></p>																																																																						

	<p>Paul, E. A., &amp; Kucey, R. M. (1981). Carbon flow in plant microbial associations. <i>Science</i>, 213(4506), 473–474.  <a href="https://doi.org/10.1126/science.213.4506.473">https://doi.org/10.1126/science.213.4506.473</a></p> <p>Rygiewicz, P. T., Miller, S. L., &amp; Durall, D. M. (1988). A root-mycocosm for growing ectomycorrhizal hyphae apart from host roots while maintaining symbiotic integrity. <i>Plant and Soil</i>, 109(2), 281–284.  <a href="https://doi.org/10.1007/bf02202096">https://doi.org/10.1007/bf02202096</a></p>
<b>Follow up Questions</b>	<p><b>Is there a CO<sub>2</sub> level where this relationship because worse for the relationship? How does this relationship affect the environment indirectly (outside of the plants and fungi)? Are specific types of Mycorrhizal fungi more responsive to CO<sub>2</sub>?</b></p>

## Article #14 Notes: How arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO<sub>2</sub>

Article notes should be on separate sheets

<b>Source Title</b>	Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO <sub>2</sub>
<b>Source citation (APA Format)</b>	<p>Cheng, L., Booker, F. L., Tu, C., Burkey, K. O., Zhou, L., Shew, H. D., Rufty, T. W., &amp; Hu, S. (2012). Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO<sub>2</sub>. <i>Science</i>, 337(6098), 1084–1087.  <a href="https://doi.org/10.1126/science.1224304">https://doi.org/10.1126/science.1224304</a></p>

<b>Original url</b>	<a href="https://doi.org/10.1126/science.1224304">https://doi.org/10.1126/science.1224304</a>
<b>Source type</b>	Article
<b>Keywords</b>	<b>Arbuscular Mycorrhizal Fungi (AMF)</b> <b>Elevated CO<sub>2</sub></b> <b>Organic Carbon Decomposition</b> <b>Soil Carbon Dynamics</b> <b>Priming Effect</b> <b>Carbon Allocation</b> <b>Photosynthate Transfer</b>
<b>#Tags</b>	#mycorrhizal networks #CO2
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- Control experiment with or without AM fungi inoculation. Plants were in growth chambers, one at ambient CO<sub>2</sub> and one at elevated CO<sub>2</sub>, isotope <sup>13</sup>C used). They measured Root colonization + hyphal length, soil respiration, and microbial biomass of soil carbon</li> <li>- Findings <ul style="list-style-type: none"> <li>○ Elevated CO<sub>2</sub> levels increased the activity of AM, leading to more colonization of plant roots and hyphal network expansion</li> <li>○ Elevated CO<sub>2</sub> levels resulted in more soil carbon losses <ul style="list-style-type: none"> <li>▪ AM stimulation under elevated CO<sub>2</sub> accelerated decomposition of C</li> </ul> </li> <li>○ Challenge the assumption that elevated CO<sub>2</sub> enhanced plant growth and mycorrhizal activity, instead it stimulated AM to accelerate C decomposition</li> </ul> </li> </ul>
<b>Research Question/Problem/Need</b>	How does elevated CO <sub>2</sub> affect AM fungi in relation to soil carbon decomposition.

<p><b>Important Figures</b></p>	<p>Effect of CO<sub>2</sub> on Nitrogen transfer within plants</p>
<p><b>VOCAB: (w/definition)</b></p>	<p>Decomposition - the state or process of rotting; decay.  Grassland - biome where grasses are the dominant vegetation  Sequestration - process by which plants remove carbon from the atmosphere through photosynthesis  Saprotrophs - organism that feeds on nonliving organic matter known as detritus at a microscopic level</p>
<p><b>Cited references to follow up on</b></p>	<p>Drigo, B., Pijl, A. S., Duyts, H., Kielak, A. M., Gamper, H. A., Houtekamer, M. J., Boschker, H. T., Bodelier, P. L., Whiteley, A. S., Veen, J. A., &amp; Kowalchuk, G. A. (2010). Shifting carbon flow from roots into associated microbial communities in response to elevated atmospheric CO<sub>2</sub>. <i>Proceedings of the National Academy of Sciences</i>, 107(24), 10938–10942.  <a href="https://doi.org/10.1073/pnas.0912421107">https://doi.org/10.1073/pnas.0912421107</a></p> <p>Wilson, G. W., Rice, C. W., Rillig, M. C., Springer, A., &amp; Hartnett, D. C. (2009). Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: Results from long-term field</p>



	<p>experiments. <i>Ecology Letters</i>, 12(5), 452–461.  <a href="https://doi.org/10.1111/j.1461-0248.2009.01303.x">https://doi.org/10.1111/j.1461-0248.2009.01303.x</a></p> <p>Hodge, A., &amp; Fitter, A. H. (2010). Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N Cycling. <i>Proceedings of the National Academy of Sciences</i>, 107(31), 13754–13759. <a href="https://doi.org/10.1073/pnas.1005874107">https://doi.org/10.1073/pnas.1005874107</a></p>
<b>Follow up Questions</b>	<b>Does C exchange benefit the plant or the fungi greater? How can soil carbon decomposition be elevated? What mechanism in AM lead to this happening?</b>

## Article #15 Notes: mycorrhizal fungi and transferring carbon between plants.

Article notes should be on separate sheets

<b>Source Title</b>	The fungus does not transfer carbon to or between roots in an arbuscular mycorrhizal symbiosis
<b>Source citation (APA Format)</b>	Pfeffer, P. E., Douds, D. D., Bücking, H., Schwartz, D. P., & Shachar-Hill, Y. (2004). The fungus does not transfer carbon to or between roots in an arbuscular mycorrhizal symbiosis. <i>New Phytologist</i> , 163(3), 617–627. <a href="https://doi.org/10.1111/j.1469-8137.2004.01152.x">https://doi.org/10.1111/j.1469-8137.2004.01152.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1111/j.1469-8137.2004.01152.x">https://doi.org/10.1111/j.1469-8137.2004.01152.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	<p><b>Arbuscular Mycorrhizal Fungi (AMF)</b></p> <p><b>Carbon Allocation</b></p> <p><b>Plant-Fungal Symbiosis</b></p> <p><b>Root Colonization</b></p> <p><b>Fungal Networks</b></p> <p><b>Carbon Flow</b></p>

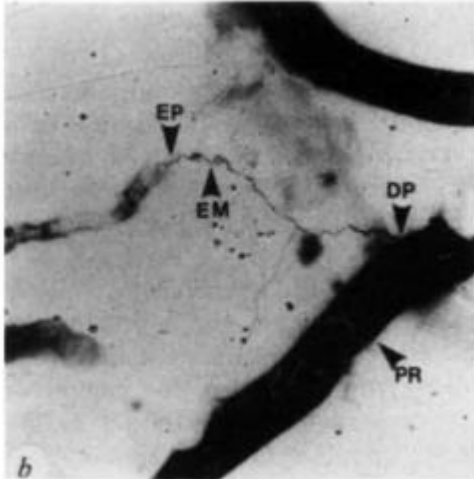
<p><b>#Tags</b></p>	<p>#mycorrhizal networks #Carbon #CMNmodel #CO2</p>																																																
<p><b>Summary of key points + notes (include methodology)</b></p>	<ul style="list-style-type: none"> <li>- Plants grown with or without fungi, connected two plants with AM relationship, plants were then given <sup>13</sup>C and isotopic analysis was used to identify carbon in fungi, soil, or plants</li> <li>- Findings             <ul style="list-style-type: none"> <li>o They were unable to find significant movement of the C between the two plants, or the fungi and the neighboring plant</li> <li>o They were unable to detect any <sup>13</sup>C in the neighboring plants, but did in the fungi</li> <li>o This shows that AM symbiosis does not do carbon transfer between plants, just uptakes for itself</li> </ul> </li> </ul>																																																
<p><b>Research Question/Problem/Need</b></p>	<p>Does AM f symbiosis facilitate transferring carbon between connected plants</p>																																																
<p><b>Important Figures</b></p>	<p>(a) Line graph showing dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt. vs time in d. Two series are plotted: one with solid circles and one with open circles. Both show an upward trend over time.</p> <table border="1"> <caption>Data for Graph (a)</caption> <thead> <tr> <th>Time (d)</th> <th>Solid Circles (dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt.)</th> <th>Open Circles (dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt.)</th> </tr> </thead> <tbody> <tr><td>7</td><td>16</td><td>1</td></tr> <tr><td>14</td><td>20</td><td>2</td></tr> <tr><td>28</td><td>22</td><td>4</td></tr> <tr><td>42</td><td>26</td><td>6</td></tr> <tr><td>56</td><td>26</td><td>8</td></tr> </tbody> </table> <p>(b) Bar graph showing % of initially added <sup>13</sup>C vs time in d. The y-axis ranges from 0 to 100. The x-axis shows time points 7, 14, 28, 42, and 56 days. The percentage decreases over time.</p> <table border="1"> <caption>Data for Graph (b)</caption> <thead> <tr> <th>Time (d)</th> <th>% of initially added <sup>13</sup>C</th> </tr> </thead> <tbody> <tr><td>7</td><td>95</td></tr> <tr><td>14</td><td>75</td></tr> <tr><td>28</td><td>48</td></tr> <tr><td>42</td><td>48</td></tr> <tr><td>56</td><td>30</td></tr> </tbody> </table> <p>(c) Bar graph showing dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt. vs time in d. The y-axis ranges from 0 to 12. The x-axis shows time points 7, 14, 28, 42, and 56 days. Two series are shown: white bars and grey bars. Both show an increase over time.</p> <table border="1"> <caption>Data for Graph (c)</caption> <thead> <tr> <th>Time (d)</th> <th>White Bars (dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt.)</th> <th>Grey Bars (dpm x 10<sup>3</sup> mg<sup>-1</sup> d. wt.)</th> </tr> </thead> <tbody> <tr><td>7</td><td>2</td><td>2.5</td></tr> <tr><td>14</td><td>7</td><td>3.5</td></tr> <tr><td>28</td><td>8</td><td>4</td></tr> <tr><td>42</td><td>4.5</td><td>9.5</td></tr> <tr><td>56</td><td>5</td><td>8</td></tr> </tbody> </table> <p>C content after roots receive glucose</p>	Time (d)	Solid Circles (dpm x 10 <sup>3</sup> mg <sup>-1</sup> d. wt.)	Open Circles (dpm x 10 <sup>3</sup> mg <sup>-1</sup> d. wt.)	7	16	1	14	20	2	28	22	4	42	26	6	56	26	8	Time (d)	% of initially added <sup>13</sup> C	7	95	14	75	28	48	42	48	56	30	Time (d)	White Bars (dpm x 10 <sup>3</sup> mg <sup>-1</sup> d. wt.)	Grey Bars (dpm x 10 <sup>3</sup> mg <sup>-1</sup> d. wt.)	7	2	2.5	14	7	3.5	28	8	4	42	4.5	9.5	56	5	8
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<p><b>VOCAB: (w/definition)</b></p>	<p>Medium - a substance that can transmit energy or light from one place to another</p> <p>fatty acids - chain-like molecules that make up fat in the body and food</p> <p>Biosynthetic metabolic pools - collective term for all of the substances involved in the metabolic process in a biological system</p>																																																

<b>Cited references to follow up on</b>	<p>Bago, B., Pfeffer, P. E., Abubaker, J., Jun, J., Allen, J. W., Brouillette, J., Douds, D. D., Lammers, P. J., &amp; Shachar-Hill, Y. (2003). Carbon export from arbuscular mycorrhizal roots involves the translocation of carbohydrate as well as lipid. <i>Plant Physiology</i>, <i>131</i>(3), 1496–1507.  <a href="https://doi.org/10.1104/pp.102.007765">https://doi.org/10.1104/pp.102.007765</a></p> <p>Bago, B., Pfeffer, P. E., Douds, D. D., Brouillette, J., Bécard, G., &amp; Shachar-Hill, Y. (1999). Carbon metabolism in spores of the arbuscular mycorrhizal fungus glomus intraradices as revealed by nuclear magnetic resonance spectroscopy. <i>Plant Physiology</i>, <i>121</i>(1), 263–272.  <a href="https://doi.org/10.1104/pp.121.1.263">https://doi.org/10.1104/pp.121.1.263</a></p> <p>Bidartondo, M. I., Redecker, D., Hijri, I., Wiemken, A., Bruns, T. D., Domínguez, L., Sérsic, A., Leake, J. R., &amp; Read, D. J. (2002). Epiparasitic plants specialized on arbuscular mycorrhizal fungi. <i>Nature</i>, <i>419</i>(6905), 389–392.  <a href="https://doi.org/10.1038/nature01054">https://doi.org/10.1038/nature01054</a></p>
<b>Follow up Questions</b>	<p><b>How does this affect the carbon symbiosis hypothesis I am researching? Does this carbon flow vary with fungi type? Could environmental conditions play a role in whether carbon is transferred or not?</b></p>

## Article #16 Notes: Transfer of carbon between plants connected with AM

Article notes should be on separate sheets

<b>Source Title</b>	Direct transfer of carbon between plants connected by vesicular–arbuscular mycorrhizal mycelium.
<b>Source citation (APA Format)</b>	Francis, R., & Read, D. J. (1984). Direct transfer of carbon between plants connected by vesicular–arbuscular

	mycorrhizal mycelium. <i>Nature</i> , 307(5946), 53–56. https://doi.org/10.1038/307053a0
Original url	<a href="https://doi.org/10.1038/307053a0">https://doi.org/10.1038/307053a0</a>
Source type	Article
Keywords	Carbon Transfer Mycorrhizal Networks Root Interconnections Plant-Fungal Symbiosis Photosynthate Allocation Isotope Labeling Common Mycorrhizal Networks (CMNs)
#Tags	#mycorrhizal networks #Carbon #CMNmodel #CO2
Summary of key points + notes (include methodology)	<ul style="list-style-type: none"> <li>- Two plants (donor and recipient) grown in same pots and colonized with fungi; control same expect no fungi. The Donor plant was given <sup>14</sup>C through CO<sub>2</sub>. Isotopic radiation was used to see how much caron transferred</li> <li>- Findings <ul style="list-style-type: none"> <li>o <sup>14</sup>C was detected in the receiving plants in the experimental not in the control</li> <li>o This shows that carbon can be transfer between plans in a network</li> </ul> </li> </ul>
Research Question/Problem/ Need	Can carbon be transferred between plants in a mycorrhizal relationship?
Important Figures	

	Picture of fungi connecting with the roots of a plant
<b>VOCAB: (w/definition)</b>	<p>Irradiation - the apparent extension of the edges of an illuminated object seen against a dark background.</p> <p>source-sink - location where resources are taken up or synthesized and used</p> <p>Inoculum - a substance used for inoculation.</p> <p>Autoradiographs - image on an X-ray film or nuclear emulsion produced by the pattern of decay emissions from a distribution of a radioactive substance</p>
<b>Cited references to follow up on</b>	<p>Heap, A. J., &amp; Newman, E. I. (1980). The influence of vesicular-arbuscular mycorrhizas on phosphorus transfer between plants. <i>New Phytologist</i>, 85(2), 173–179.  <a href="https://doi.org/10.1111/j.1469-8137.1980.tb04458.x">https://doi.org/10.1111/j.1469-8137.1980.tb04458.x</a></p> <p>Chiariello, N., Hickman, J. C., &amp; Mooney, H. A. (1982). Endomycorrhizal role for interspecific transfer of phosphorus in a community of annual plants. <i>Science</i>, 217(4563), 941–943.  <a href="https://doi.org/10.1126/science.217.4563.941">https://doi.org/10.1126/science.217.4563.941</a></p> <p>Cavagnaro, T., Gao, L., Smith, F., &amp; Smith, S. (2001). Morphology of arbuscular mycorrhizas is influenced by fungal identity. <i>New Phytologist</i>, 151(2), 469–475.  <a href="https://doi.org/10.1046/j.0028-646x.2001.00191.x">https://doi.org/10.1046/j.0028-646x.2001.00191.x</a></p>
<b>Follow up Questions</b>	<b>What factors affect the magnitude and direction of the transfer? Does carbon affect the recipient plant? How does carbon transfer differ from other molecular transfer?</b>

## Article #17 Notes: effects of CO<sub>2</sub> on mycorrhizal colonization

Article notes should be on separate sheets

<b>Source Title</b>	Effect of elevated atmospheric CO <sub>2</sub> on mycorrhizal colonization, external mycorrhizal hyphal production and phosphorus inflow in <i>Plantago lanceolata</i> and <i>Trifolium repens</i> in association with the arbuscular mycorrhizal fungus <i>Glomus mosseae</i>
<b>Source citation (APA Format)</b>	Staddon, P. L., Fitter, A. H., & Graves, J. D. (1999). Effect of elevated atmospheric CO <sub>2</sub> on mycorrhizal colonization, external mycorrhizal hyphal production and phosphorus inflow in <i>plantago lanceolata</i> and <i>trifolium repens</i> in association with the arbuscular mycorrhizal fungus <i>Glomus Mosseae</i> . <i>Global Change Biology</i> , 5(3), 347–358. <a href="https://doi.org/10.1046/j.1365-2486.1999.00230.x">https://doi.org/10.1046/j.1365-2486.1999.00230.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1046/j.1365-2486.1999.00230.x">https://doi.org/10.1046/j.1365-2486.1999.00230.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	<b>Elevated CO<sub>2</sub></b> <b>Arbuscular Mycorrhizal Fungi (AMF)</b> <b>Glomus mosseae</b> <b>Mycorrhizal Colonization</b> <b>External Hyphal Production</b> <b>Phosphorus Inflow</b>
<b>#Tags</b>	#mycorrhizal networks #Carbon #CMNmodel #CO <sub>2</sub> #CO <sub>2</sub> change
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- Plants <i>Plantago lanceolata</i> (ribwort plantain) and <i>Trifolium repens</i> (white clover) inoculated with <i>Glomus mosseae</i> to establish relationship. Then two conditions of CO<sub>2</sub> exposed too Ambient CO<sub>2</sub> concentration (~350 ppm) and elevated CO<sub>2</sub> concentration (~700 ppm), simulating predicted future scenarios. phosphorus was then put in the soil (uptake measured as well as root and soil samples)</li> <li>- Findings <ul style="list-style-type: none"> <li>o Elevated CO<sub>2</sub> led to increase in % of root length of AM fungus and higher density of the hyphae, however data analysis led to</li> </ul> </li> </ul>

	<p>find this data to be insignificant and do to plant size then direct CO2 effect</p> <ul style="list-style-type: none"> <li>○ Root phosphorous remained unchanged</li> <li>○ CO2 promotes plant growth but does not affect the relationship between the fungi and the plants.</li> </ul>																																								
<p><b>Research Question/Problem/Need</b></p>	<p>How does CO2 affect the symbiotic relationship between AM fungi and host plants?</p>																																								
<p><b>Important Figures</b></p>	<table border="1" data-bbox="643 569 1401 1041"> <thead> <tr> <th colspan="4" style="background-color: #e0e0e0;">elevated CO<sub>2</sub> treatment effect</th> </tr> <tr> <th style="background-color: #e0e0e0;"></th> <th style="background-color: #e0e0e0;">Covariate</th> <th style="background-color: #e0e0e0;">p-value</th> <th style="background-color: #e0e0e0;">Direction</th> </tr> </thead> <tbody> <tr> <td>Total plant DW</td> <td>age</td> <td>**</td> <td>increase</td> </tr> <tr> <td>Shoot DW</td> <td>age</td> <td>**</td> <td>increase</td> </tr> <tr> <td>Root DW</td> <td>age</td> <td>***</td> <td>increase</td> </tr> <tr> <td>Root length</td> <td>age</td> <td>**</td> <td>increase</td> </tr> <tr> <td>Specific root length (SRL)</td> <td>age</td> <td>NS</td> <td>NA</td> </tr> <tr> <td>SRL</td> <td>total plant DW</td> <td>NS</td> <td>NA</td> </tr> <tr> <td>Root DW</td> <td>shoot DW</td> <td>NS</td> <td>NA</td> </tr> <tr> <td>Root DW</td> <td>total plant DW</td> <td>NS</td> <td>NA</td> </tr> </tbody> </table> <p>CO2 effect on plant growth</p>	elevated CO <sub>2</sub> treatment effect					Covariate	p-value	Direction	Total plant DW	age	**	increase	Shoot DW	age	**	increase	Root DW	age	***	increase	Root length	age	**	increase	Specific root length (SRL)	age	NS	NA	SRL	total plant DW	NS	NA	Root DW	shoot DW	NS	NA	Root DW	total plant DW	NS	NA
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<p><b>VOCAB: (w/definition)</b></p>	<p>Covariate - a variable that is related to a dependent variable and can be used to predict or explain it</p> <p>Nonparametric - a type of statistical analysis that makes few or no assumptions about the distribution of the data being studied</p> <p>Sequential - forming or following in a logical order or sequence</p> <p>air turbulence - when air flows in a chaotic or random way, rather than smoothly in one direction</p> <p>gas analyzer - scientific devices that measure the concentration of a particular gas in a mixture of multiple gases</p>																																								
<p><b>Cited references to follow up on</b></p>	<p>Emery, S. M., Bell-Dereske, L., Stahlheber, K. A., &amp; Gross, K. L. (2022a). Arbuscular mycorrhizal fungal community responses to drought and nitrogen fertilization in switchgrass stands. <i>Applied Soil Ecology</i>, 169, 104218. <a href="https://doi.org/10.1016/j.apsoil.2021.104218">https://doi.org/10.1016/j.apsoil.2021.104218</a></p>																																								

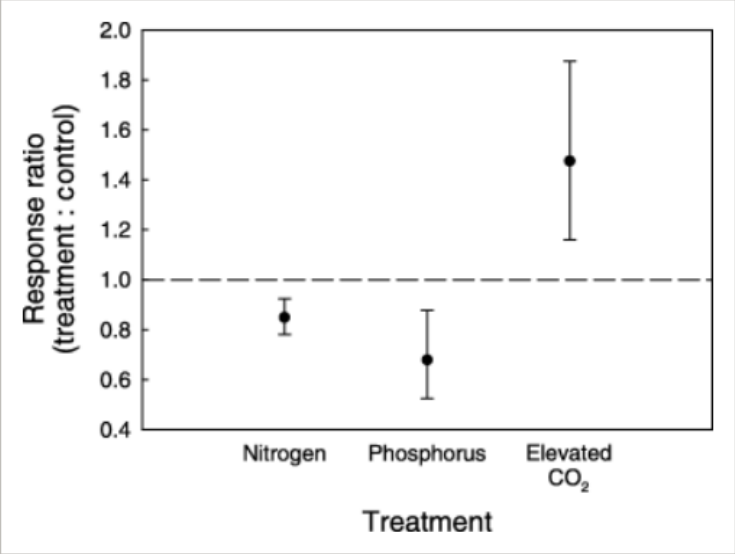
<b>Follow up Questions</b>	<b>What was the difference identified between the two plant types? What would account for this? What plants are best when experimenting on nutrient transfer with AM fungi?</b>
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## Article #18 Notes: Mycorrhizal response to nutrients and CO<sub>2</sub> variation

Article notes should be on separate sheets

<b>Source Title</b>	A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric CO <sub>2</sub> in field studies
<b>Source citation (APA Format)</b>	Treseder, K. K. (2004). A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric CO <sub>2</sub> in field studies. <i>New Phytologist</i> , 164(2), 347–355. <a href="https://doi.org/10.1111/j.1469-8137.2004.01159.x">https://doi.org/10.1111/j.1469-8137.2004.01159.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1111/j.1469-8137.2004.01159.x">https://doi.org/10.1111/j.1469-8137.2004.01159.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	<b>Meta-Analysis</b> <b>Mycorrhizal Fungi</b> <b>Nitrogen Deposition</b> <b>Phosphorus Addition</b> <b>Elevated CO<sub>2</sub></b> <b>Field Studies</b> <b>Nutrient Cycling</b> <b>Plant-Fungal Symbiosis</b>
<b>#Tags</b>	#mycorrhizal networks #Carbon #CMNmodel #CO <sub>2</sub> #N #P
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- Conducted literary search and analysis to examine the effects of N and P additions as well as elevated CO<sub>2</sub> on the AM fungi</li> <li>- Findings <ul style="list-style-type: none"> <li>○ Nitrogen additions</li> </ul> </li> </ul>



	<ul style="list-style-type: none"> <li>▪ Fungi abundance decreased by 15% following nitrogen addition</li> <li>▪ Suggests increasing nitrogen availability may reduce plants reliance on mycorrhizal relationship for nutrients</li> <li>○ Phosphorus audients             <ul style="list-style-type: none"> <li>▪ No significant effect found</li> <li>▪ Not a limiting factor on fungi</li> </ul> </li> <li>○ Elevated CO2             <ul style="list-style-type: none"> <li>▪ Average increase of 47% in abundance for elevated CO2</li> <li>▪ May enhance plant carbon allocations, strengthening symbiotic relationships.</li> </ul> </li> </ul>								
<p><b>Research Question/Problem/Need</b></p>	<p>How do different factors (N, P, and elevated CO2) affect mycorrhizal abundance?</p>								
<p><b>Important Figures</b></p>	<div style="text-align: center;">  <table border="1" style="margin-left: auto; margin-right: auto;"> <caption>Response Ratio Data</caption> <thead> <tr> <th>Treatment</th> <th>Response Ratio (approx.)</th> </tr> </thead> <tbody> <tr> <td>Nitrogen</td> <td>0.85</td> </tr> <tr> <td>Phosphorus</td> <td>0.7</td> </tr> <tr> <td>Elevated CO<sub>2</sub></td> <td>1.45</td> </tr> </tbody> </table> </div> <p>Response of plants in relationship compared to that with CO2 and ones without</p>	Treatment	Response Ratio (approx.)	Nitrogen	0.85	Phosphorus	0.7	Elevated CO <sub>2</sub>	1.45
Treatment	Response Ratio (approx.)								
Nitrogen	0.85								
Phosphorus	0.7								
Elevated CO <sub>2</sub>	1.45								
<p><b>VOCAB: (w/definition)</b></p>	<p>Sporocarps - structures that produce and release spores in a variety of organisms, including fungi, aquatic ferns, and amoebozoans</p> <p>Abundance - a very large quantity of something</p> <p>meta-analyses - a statistical method that combines data from multiple studies to draw conclusions about a common research question</p>								

	<p>Omitted - leave out or exclude (someone or something), either intentionally or forgetfully</p> <p>Nitrate - a compound of nitrogen and oxygen naturally found in air, soil, water, and some food</p>
<b>Cited references to follow up on</b>	<p>Cornwell, W. K., Bedford, B. L., &amp; Chapin, C. T. (2001). Occurrence of arbuscular mycorrhizal fungi in a phosphorus-poor wetland and mycorrhizal response to phosphorus fertilization. <i>American Journal of Botany</i>, 88(10), 1824–1829. <a href="https://doi.org/10.2307/3558359">https://doi.org/10.2307/3558359</a></p> <p>Lukac, M., Caleapietra, C., &amp; Godbold, D. L. (2003). Production, turnover and mycorrhizal colonization of root systems of three populus species grown under elevated co2 . <i>Global Change Biology</i>, 9(6), 838–848. <a href="https://doi.org/10.1046/j.1365-2486.2003.00582.x">https://doi.org/10.1046/j.1365-2486.2003.00582.x</a></p> <p>Poorter, H. (1993). Interspecific variation in the growth response of plants to an elevated ambient CO2 concentration. <i>CO2 and Biosphere</i>, 77–98. <a href="https://doi.org/10.1007/978-94-011-1797-5_6">https://doi.org/10.1007/978-94-011-1797-5_6</a></p>
<b>Follow up Questions</b>	<b>Why does N reduce abundance? Is it only do to plants having access to N on its own? Why is this effect not</b>

## Article #19 Notes: Fungi and plant response to CO2 increase

Article notes should be on separate sheets

<b>Source Title</b>	Taking mycoentrism seriously: Mycorrhizal fungal and plant responses to elevated co2
<b>Source citation (APA Format)</b>	Alberton, O., Kuyper, T. W., & Gorissen, A. (2005). Taking mycoentrism seriously: Mycorrhizal fungal and plant

	responses to elevated co2. <i>New Phytologist</i> , 167(3), 859–868. <a href="https://doi.org/10.1111/j.1469-8137.2005.01458.x">https://doi.org/10.1111/j.1469-8137.2005.01458.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1111/j.1469-8137.2005.01458.x">https://doi.org/10.1111/j.1469-8137.2005.01458.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	<p><b>Mycocentrism</b></p> <p><b>Arbuscular Mycorrhizal Fungi (AMF)</b></p> <p><b>Ectomycorrhizal Fungi (ECM)</b></p> <p><b>Elevated CO<sub>2</sub></b></p> <p><b>Carbon Allocation</b></p> <p><b>Fungal Abundance</b></p> <p><b>Plant-Fungal Symbiosis</b></p>
<b>#Tags</b>	#mycorrhizal networks #Carbon #CMNmodel #CO2 #CO2change
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- They put plants in a relationship with both ECM and AM they conducted experiments under elevated CO<sub>2</sub> levels and ambient to simulate future atmospheric conditions</li> <li>- They then measured for fungal colonization, biomass, and hyphal and plant growth as well as isotopic labeling <sup>13</sup>C for movement of carbon</li> <li>- Findings <ul style="list-style-type: none"> <li>○ Elevated CO<sub>2</sub> resulted in a significant increase in abundance.</li> <li>○ ECM fungi exhibited a 34% increase in abundance, AM fungi showed a 21% increase <ul style="list-style-type: none"> <li>▪ bigger response in ECM fungi</li> </ul> </li> <li>○ No significant difference in growth</li> <li>○ ECM found monocentric perspective, but not in AM <ul style="list-style-type: none"> <li>▪ ECM and AM have different dynamics related to carbon</li> </ul> </li> <li>○ Elevated CO<sub>2</sub> may alter the balance of carbon allocation + nutrient transfer differently in ECM and AM networks</li> </ul> </li> </ul>
<b>Research Question/Problem/Need</b>	How do AM fungi and ECM fungi react to elevated CO <sub>2</sub> with abundance and activity?

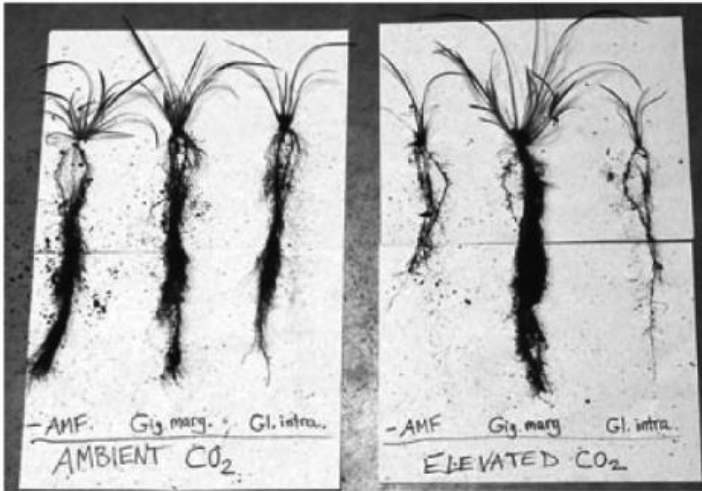
<p><b>Important Figures</b></p>	<table border="1"> <thead> <tr> <th>Categories</th> <th>R</th> <th>95% CI</th> <th>n</th> <th>P</th> </tr> </thead> <tbody> <tr> <td>AM plants</td> <td>1.25</td> <td>1.19–1.31</td> <td>74</td> <td></td> </tr> <tr> <td>ECM fungi</td> <td>1.34</td> <td>1.25–1.45</td> <td>65</td> <td>0.17</td> </tr> <tr> <td>ECM plants</td> <td>1.26</td> <td>1.19–1.34</td> <td>65</td> <td></td> </tr> <tr> <td>AM fungi</td> <td>1.21</td> <td>1.12–1.32</td> <td>77</td> <td>0.32</td> </tr> <tr> <td>AM plants</td> <td>1.25</td> <td>1.19–1.31</td> <td>74</td> <td></td> </tr> <tr> <td>ECM extraradical</td> <td>1.45</td> <td>1.30–1.65</td> <td>38</td> <td>&lt; 0.01</td> </tr> <tr> <td>ECM colonization percentage</td> <td>1.19</td> <td>1.09–1.28</td> <td>31</td> <td></td> </tr> <tr> <td>AM extraradical</td> <td>1.23</td> <td>1.07–1.40</td> <td>46</td> <td>0.65</td> </tr> <tr> <td>AM colonization percentage</td> <td>1.17</td> <td>1.06–1.30</td> <td>30</td> <td></td> </tr> <tr> <td>ECM extraradical</td> <td>1.45</td> <td>1.30–1.65</td> <td>38</td> <td>0.04</td> </tr> <tr> <td>AM extraradical</td> <td>1.23</td> <td>1.07–1.40</td> <td>46</td> <td></td> </tr> <tr> <td>ECM extraradical</td> <td>1.45</td> <td>1.30–1.65</td> <td>38</td> <td>0.03</td> </tr> <tr> <td>ECM plants</td> <td>1.26</td> <td>1.19–1.34</td> <td>65</td> <td></td> </tr> <tr> <td>AM plants</td> <td>1.25</td> <td>1.19–1.31</td> <td>74</td> <td>0.55</td> </tr> <tr> <td>AM extraradical</td> <td>1.23</td> <td>1.07–1.40</td> <td>46</td> <td></td> </tr> </tbody> </table> <p>Meta-analysis of the effects of elevated CO<sub>2</sub> on mycorrhizal systems</p>	Categories	R	95% CI	n	P	AM plants	1.25	1.19–1.31	74		ECM fungi	1.34	1.25–1.45	65	0.17	ECM plants	1.26	1.19–1.34	65		AM fungi	1.21	1.12–1.32	77	0.32	AM plants	1.25	1.19–1.31	74		ECM extraradical	1.45	1.30–1.65	38	< 0.01	ECM colonization percentage	1.19	1.09–1.28	31		AM extraradical	1.23	1.07–1.40	46	0.65	AM colonization percentage	1.17	1.06–1.30	30		ECM extraradical	1.45	1.30–1.65	38	0.04	AM extraradical	1.23	1.07–1.40	46		ECM extraradical	1.45	1.30–1.65	38	0.03	ECM plants	1.26	1.19–1.34	65		AM plants	1.25	1.19–1.31	74	0.55	AM extraradical	1.23	1.07–1.40	46	
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<p><b>VOCAB: (w/definition)</b></p>	<p>meta-analysis - a statistical process that combines data from multiple studies to draw conclusions about a topic</p> <p>Ozone - gas made up of three oxygen atoms</p> <p>PLFA - phospholipid fatty acids</p> <p>extraradical mycelium - a network of fungal hyphae that extends from the roots of plants into the soil</p>																																																																																
<p><b>Cited references to follow up on</b></p>	<p>Gavito, M. E. (2000). Atmospheric CO<sub>2</sub> and mycorrhiza effects on biomass allocation and nutrient uptake of nodulated pea (<i>Pisum sativum</i> L.) plants. <i>Journal of Experimental Botany</i>, 51(352), 1931–1938.  <a href="https://doi.org/10.1093/jexbot/51.352.1931">https://doi.org/10.1093/jexbot/51.352.1931</a></p> <p>Hu, S., Wu, J., Burkey, K. O., &amp; Firestone, M. K. (2005). Plant and Microbial N acquisition under elevated atmospheric CO<sub>2</sub> in two mesocosm experiments with annual grasses. <i>Global Change Biology</i>, 11(2), 213–223.  <a href="https://doi.org/10.1111/j.1365-2486.2005.00905.x">https://doi.org/10.1111/j.1365-2486.2005.00905.x</a></p>																																																																																

	Lussenhop, J., Treonis, A., Curtis, P. S., Teeri, J. A., & Vogel, C. S. (1998). Response of soil biota to elevated atmospheric CO <sub>2</sub> in Poplar Model Systems. <i>Oecologia</i> , 113(2), 247–251. <a href="https://doi.org/10.1007/s004420050375">https://doi.org/10.1007/s004420050375</a>
<b>Follow up Questions</b>	<b>Why do ECM fungi have a stronger response to elevated CO<sub>2</sub>? How does increased fungal abundance under the elevated CO<sub>2</sub> affect the fungi's nutrient uptake? Is using ECM more beneficial than AM?</b>

## Article #20 Notes: plants associated with working with mycorrhizal fungi and CO<sub>2</sub> increase

Article notes should be on separate sheets

<b>Source Title</b>	Species of plants and associated arbuscular mycorrhizal fungi mediate mycorrhizal responses to CO <sub>2</sub> enrichment
<b>Source citation (APA Format)</b>	Johnson, N. C., Wolf, J., Reyes, M. A., Panter, A., Koch, G. W., & Redman, A. (2005). Species of plants and associated arbuscular mycorrhizal fungi mediate mycorrhizal responses to CO <sub>2</sub> enrichment. <i>Global Change Biology</i> , 11(7), 1156–1166. <a href="https://doi.org/10.1111/j.1365-2486.2005.00967.x">https://doi.org/10.1111/j.1365-2486.2005.00967.x</a>
<b>Original url</b>	<a href="https://doi.org/10.1111/j.1365-2486.2005.00967.x">https://doi.org/10.1111/j.1365-2486.2005.00967.x</a>
<b>Source type</b>	Article
<b>Keywords</b>	Arbuscular Mycorrhizal Fungi (AMF) Symbiotic Relationships Elevated CO <sub>2</sub> Plant-Fungal Interactions Carbon Allocation

	<b>Nutrient Uptake</b>
<b>#Tags</b>	#mycorrhizal networks #Carbon #CMNmodel #CO2 #CO2change
<b>Summary of key points + notes (include methodology)</b>	<ul style="list-style-type: none"> <li>- Plants were inoculated in pairing, with multiple different species used. They were then grown under ambient CO<sub>2</sub> (current atmospheric levels) and elevated CO<sub>2</sub> (predicted levels) They then measured plant growth and <sup>13</sup>C allocation between test and control</li> <li>- Findings <ul style="list-style-type: none"> <li>o Different plants exhibited different responses to the fungi</li> <li>o This shows that plant chosen matters</li> <li>o CO<sub>2</sub> increased the nutrient uptake amounts (it therefore enhances mutualistic benefits of the fungi)</li> </ul> </li> </ul>
<b>Research Question/Problem/Need</b>	How do specific plants react to AM fungal relationship?
<b>Important Figures</b>	 <p>Root length of plants grown with different fungi in different CO<sub>2</sub> conditions</p>
<b>VOCAB: (w/definition)</b>	<ul style="list-style-type: none"> <li>- Species-Specific Effects: The unique responses and interactions that occur based on the identity of plant and fungal species in a symbiotic relationship.</li> <li>- Symbiotic Specificity: The degree to which a particular plant and fungal species form an exclusive or preferential symbiotic relationship.</li> </ul>

	<ul style="list-style-type: none"> <li>- Hyphal Network: The extensive web of fungal filaments (hyphae) in the soil that connects plant roots and facilitates the transfer of nutrients and carbon.</li> <li>- Mutualism-Dependency Gradient: A continuum describing the degree to which plants rely on mycorrhizal fungi for nutrient acquisition, ranging from high mutualism (mutually beneficial) to low dependency (minimal reliance).</li> </ul>
<b>Cited references to follow up on</b>	<p>Diaz, S. (1995). Elevated co<sub>2</sub> responsiveness, interactions at the community level and plant functional types. <i>Journal of Biogeography</i>, 22(2/3), 289.  <a href="https://doi.org/10.2307/2845923">https://doi.org/10.2307/2845923</a></p> <p>Fitter, A. H., Heinemeyer, A., &amp; Staddon, P. L. (2000). The impact of Elevated Co<sub>2</sub> and global climate change on Arbuscular Mycorrhizas: A mycocentric approach. <i>New Phytologist</i>, 147(1), 179–187.  <a href="https://doi.org/10.1046/j.1469-8137.2000.00680.x">https://doi.org/10.1046/j.1469-8137.2000.00680.x</a></p> <p>Gavito, Mayra E, Schweiger, P., &amp; Jakobsen, I. (2002). P uptake by arbuscular mycorrhizal hyphae: Effect of soil temperature and atmospheric CO<sub>2</sub> enrichment. <i>Global Change Biology</i>, 9(1), 106–116.  <a href="https://doi.org/10.1046/j.1365-2486.2003.00560.x">https://doi.org/10.1046/j.1365-2486.2003.00560.x</a></p>
<b>Follow up Questions</b>	<p><b>Why do different plants react differently in the network? How do the fungi specifically benefit from receiving more carbon? Could environmental factors influence the relationship in this study?</b></p>